

Identification and fungicide sensitivity of *Microdochium chrysopogonis* (Ascomycota, Amphisphaeriaceae), a new species causing tar spot of *Chrysopogon zizanioides* in southern China

Xiang Lu¹, Mengxian Mai¹, Wenhui Tan¹, Muyan Zhang¹, Jie Xie¹, Yi Lu¹, Xue Li Niu¹, Wu Zhang²

¹ School of Life Sciences and Technology, Lingnan Normal University, Zhanjiang 524048, China

² School of Geographical Science, Lingnan Normal University, Zhanjiang 524048, China

Corresponding author: Wu Zhang (ldzw1987@163.com)

Abstract

Vetiver grass (*Chrysopogon zizanioides*) has received extensive attention in recent years due to its diverse applications in soil and water conservation, heavy metal remediation, as well as essential oil and phenolic acids extraction. In 2019, the emergence of tar spot disease on *C. zizanioides* was documented in Zhanjiang, Guangdong Province, China. Initially, the disease manifested as black ascomata embedded within leaf tissue, either scattered or clustered on leaf surfaces. Subsequently, these ascomata became surrounded by fisheye lesions, characterised by brown, elliptical, necrotic haloes, which eventually coalesced, resulting in leaf withering. Koch's postulates demonstrated that the fungus isolated from these lesions was the causal agent. Microscopic examination showed that the pathogen morphologically belonged to *Microdochium*. The phylogenetic tree inferred from the combined ITS, LSU, *tub2* and *rpb2* sequences revealed the three isolates including GDMCC 3.683, LNU-196 and LNU-197 to be a novel species of *Microdochium*. Combining the results of phylogenetic, pathogenicity and morphological analyses, we propose a new species named *M. chrysopogonis* as the causal agent of *C. zizanioides* in southern China. The optimum growth temperature for *M. chrysopogonis* was determined to be 30 °C. The in vitro fungicide sensitivity of *M. chrysopogonis* was determined using a mycelial growth assay. Four demethylation-inhibiting (DMI) fungicides, including difenoconazole, flusilazole, propiconazole and tebuconazole and one methyl benzimidazole carbamate (MBC) fungicide, carbendazim, were effective against *M. chrysopogonis*, with mean 50% effective concentration (EC₅₀) values of 0.077, 0.011, 0.004, 0.024 and 0.007 µg/ml, respectively. These findings provide essential references for the precise diagnosis and effective management of *M. chrysopogonis*.

Key words: fungicide sensitivity, multilocus phylogeny, new taxon, pathogenicity, tar spot



Academic editor: C. Phukhamsakda

Received: 7 September 2023

Accepted: 19 November 2023

Published: 6 December 2023

Citation: Lu X, Mai M, Tan W, Zhang M, Xie J, Lu Y, Niu XL, Zhang W (2023) Identification and fungicide sensitivity of *Microdochium chrysopogonis* (Ascomycota, Amphisphaeriaceae), a new species causing tar spot of *Chrysopogon zizanioides* in southern China. MycoKeys 100: 205–232. <https://doi.org/10.3897/mycokeys.100.112128>

Copyright: © Xiang Lu et al.

This is an open access article distributed under terms of the Creative Commons Attribution License (Attribution 4.0 International – CC BY 4.0).

Introduction

Vetiver (*Chrysopogon zizanioides*) is one of the main grasses in tropical and subtropical areas (Maurya et al. 2023). With a large root system that penetrates deep into the soil, *C. zizanioides* is strongly tolerant to adverse environments, such as drought, salinity and heavy metals. Recently, vetiver has been widely

utilised in various applications, such as land restoration, soil and water conservation and phytoremediation of heavy metal-contaminated soils (Chen et al. 2021). Moreover, the essential oil and phenolic acids extracted from vetiver root possess significant aromatic and biological properties, giving it an important role in perfumery, the food industry and medicine (David et al. 2019; Moon et al. 2020).

To our knowledge, four diseases on *C. zizanioides* have been reported, namely, leaf blight caused by *Curvularia trifolii* in India (Babu et al. 2021), root and basal stem rot caused by *Gaeumannomyces graminicola* (Lin et al. 2019), leaf spot caused by *Phoma herbarum* (Zhang et al. 2017) and leaf streak caused by *Stenocarpella chrysopogonis* (Jia et al. 2023).

Microdochium species were originally introduced with the type species, *M. phragmitis*, identified on the leaves of *Phragmites australis* in Germany (Sydow 1924). Presently, there are 49 species included in this genus. However, only a subset of these species can induce diseases, primarily affecting grasses and cereals. For example, *M. albescens* (also referred to as *Monographella albescens*) typically induces leaf scald and grain discoloration in rice, leading to a global reduction in rice yield (Araujo et al. 2016; Dirchwolf et al. 2023). *M. bolleyi* is recognised for inducing root necrosis and basal rot in creeping bent grass in Korea, as well as causing root rot on triticale in Kazakhstan (Hong et al. 2008; Alkan et al. 2021). *M. nivale* and *M. majus* frequently result in the occurrence of pink snow mould or Fusarium Patch on wheat, barley and turf grass in cold to temperate regions (Ren et al. 2015; Abdelhalim et al. 2020). *M. opuntiae* leads to brown spotting on *Opuntia* (Braun 1995). *M. poae* triggers leaf blight disease in turf-grasses, such as *Poa pratensis* and *Agrostis stolonifera* (Liang et al. 2019). *M. paspali* is recognised for its ability to cause leaf blight in seashore paspalum (*Paspalum vaginatum*) (Zhang et al. 2015). *M. panattonianum* has the potential to induce anthracnose in lettuce (Galea et al. 1986). *M. sorghi* is accountable for the formation of zonate leaf spots and decay on sorghum species (Stewart et al. 2019).

The application of fungicides has always been an effective approach for disease control. In recent decades, demethylation-inhibiting (DMI) fungicides have emerged as a significant and extensive group of fungicides, exhibiting notable efficacy in the control of diseases caused by the *Microdochium* genus. Notably, compounds such as prochloraz, difenoconazole, propiconazole, metconazole, myclobutanil, tebuconazole and triticonazole have shown substantial antifungal efficacy against *M. panattonianum*, *M. majus* and *M. nivale* (Wicks et al. 1994; Debieu et al. 2000; Glynn et al. 2008; Mao et al. 2023). Additionally, fungicide subgroups, including phenylpyrrole (PP) fungicides, such as fludioxonil, dicarboximides, such as iprodione and quinone outside inhibitors (Qols), such as trifloxystrobin, have demonstrated noteworthy efficacy in the management of diseases induced by *M. nivale* (Glynn et al. 2008; Koch et al. 2015; Aamlid et al. 2017). Therefore, to promote effective control against tar spot of *C. zizanioides*, it is necessary to determine the sensitivity of the pathogen to fungicides.

The main objectives of this study were to identify the pathogenic fungi causing tar spot of *C. zizanioides* in southern China on the basis of morphological characteristics and multigene sequence analysis; to determine the pathogenicity to *C. zizanioides*; and to determine the inhibitory effect of fungicides against mycelial growth of the pathogen.

Materials and methods

Sample collection and fungal isolation

Leaves exhibiting symptoms of tar spot on *C. zizanioides* were collected in fields of the Grass Research Station of Lingnan Normal University (**LNU**), Zhanjiang, Guangdong, China. Leaf segments (0.5 × 0.5 cm) from the transition zone from diseased to healthy tissue were cut and surface-sterilised for 30 s with 75% ethanol and 2% sodium hypochlorite (NaClO) for 1 min, rinsed with distilled water 3 times, dried on sterile filter paper and placed on 2% potato dextrose agar (PDA) (Crous et al. 2021b). Additionally, ascomata developing on the surface of diseased tissue were gently scraped using a sterile scalpel. Subsequently, a small number of ascospores were transferred and evenly spread on to the surface of a water agar (WA) plate. Hyphal tips originating from leaf tissue fragments and single germinating conidia were transferred on to PDA medium. They were then incubated at 30 °C in darkness (Polizzi et al. 2009). After a period of 7 days, the isolates were transferred on to PDA slants and preserved at 4 °C in the culture collection of Lingnan Normal University. Additionally, they were deposited in the Guangdong Microbial Culture Collection Center (**GDMCC**) in Guangzhou, China. The holotype specimen was preserved in the Herbarium of the Chinese Academy of Forestry (**CAF**) in Beijing, China.

Morphological characterisation

Colonies were subcultured on 2% malt extract agar (MEA) and oatmeal agar (OA) at 30 °C for 10 days in the dark (Crous et al. 2009). Colony colour was characterised using Rayner's Mycological Color Chart (Rayner 1970) and colony diameters were measured after incubation for 10 days at 30 °C in the dark. Morphological characters of ascomata, asci, ascospores, sporodochia, hyphae, conidiomata, conidiophores, conidiogenous cells and conidia were determined in sterile water using an Olympus BX53 compound microscope (Tokyo, Japan), equipped with cellSens Dimension software (version 1.17).

DNA extraction, PCR amplification and sequencing

Fungal genomic DNA was extracted from mycelia grown on PDA medium after 10 days using the ENZA Fungal DNA Miniprep Kit (Omega Bio-tek, Doraville, Norcross, GA, U.S.A.), according to the protocol of manufacturer. Four loci, including internal transcribed spacer (ITS) rDNA region, large subunit ribosomal acid (LSU) rDNA region, RNA polymerase II second largest subunit gene (*rpb2*) and part of the beta-tubulin gene (*tub2*), were amplified by the following primer pairs: ITS1 and ITS4 for ITS (White et al. 1990), LR0R and LR5 for LSU (Vilgalys and Hester 1990), RPB150F (Jewell and Hsiang 2013) and fRPB2-7Cr (Liu et al. 1999) and Btub526F and Btub1332R (Jewell and Hsiang 2013). The polymerase chain reaction (PCR) conditions were as follows: 94 °C for 5 min; 94 °C for 30 s, annealing temperature for 45 s and 72 °C for 1 min, 35 cycles; and a final extension step at 72 °C for 10 min. The annealing temperature for ITS and LSU was 54 °C, for *tub2* was 55 °C and for *rpb2* was 57 °C. PCR products were sequenced by Sangon Biotech Co., Ltd. (Shanghai, China). Sequences were edited and assembled using DNAMAN version 5.2.2 and deposited in the NCBI GenBank nucleotide database (Table 1).

Table 1. Strains included in the phylogenetic analyses with collection details and GenBank accession numbers.

Species	Voucher	Country	GenBank Accession Number			
			LSU	ITS	<i>tub2</i>	<i>rpb2</i>
<i>Microdochium albescens</i>	CBS 290.79	Ivory Coast	KP858950	KP859014	KP859078	KP859123
	CBS 291.79	Ivory Coast	KP858932	KP858996	KP859059	KP859105
	CBS 243.83	Unknown country	KP858930	KP858994	KP859057	KP859103
<i>M. bolleyi</i>	CBS 172.63	Germany	MH869857	MH858255	–	–
	CBS 540.92	Syria	KP858946	KP859010	KP859073	KP859119
	Kaz_Mb01	Kazakhstan	–	MW301448	–	–
	Kaz_Mb02	Kazakhstan	–	MW301449	–	–
	CBS 137.64	Netherlands	MH870023	MH858394	–	–
	CPC 25994	Canada	KP858954	KP859018	KP859074	KP859127
	CBS 102891	Germany	MH874405	–	–	–
	CBS 618.72	Germany	MH872294	MH860598	–	–
<i>M. chrysanthemoides</i>	CGMCC 3.17929 ^T	China	KU746736	KU746690	KU746781	–
	CGMCC 3.17930	China	KU746735	KU746689	KU746782	–
<i>M. chuxiongense</i>	YFCC 8794 ^T	China	OK586160	OK586161	OK556901	OK584019
<i>M. citrinidiscum</i>	CBS 109067 ^T	Peru	KP858939	KP859003	KP859066	KP859112
<i>M. colombiense</i>	CBS 624.94 ^T	Colombia	KP858935	KP858999	KP859062	KP859108
<i>M. dawsoniorum</i>	BRIP 65649 ^T	Australia	ON394569	MK966337	–	–
	BRIP 67439	Australia	OM333563	MN492650	–	ON624208
<i>M. fisheri</i>	CBS 242.90 ^T	UK	KP858951	KP859015	KP859079	KP859124
	NFCCI 4083	India	KY777594	KY777595	–	–
	C30 ITI	Sri Lanka	–	MT875317	–	–
<i>M. graminearum</i>	CGMCC 3.23525 ^T	China	OP104016	OP103966	OP236029	OP236026
	CGMCC 3.23524	China	OP104015	OP103965	OP242835	OP236026
<i>M. hainanense</i>	SAUCC210781 ^T	China	OM959323	OM956295	OM981146	OM981153
	SAUCC210782	China	OM959324	OM956296	OM981147	OM981154
<i>M. indocalami</i>	SAUCC1016 ^T	China	MT199878	MT199884	MT435653	MT510550
<i>M. insulare</i>	BRIP 75114a	Australia	OQ892168	OQ917075	–	OQ889560
<i>M. lycopodium</i>	CBS 146.68	The Netherlands	KP858929	KP858993	KP859056	KP859102
	CBS 109397	Germany	KP858940	KP859004	KP859067	KP859113
	CBS 109398	Germany	KP858941	KP859005	KP859068	KP859114
	CBS 109399	Germany	KP858942	KP859006	KP859069	KP859115
	CBS 125585 ^T	Austria	KP858952	KP859016	KP859080	KP859125
<i>M. maculosum</i>	COAD 3358 ^T	Brazil	OK966953	OK966954	–	OL310501
<i>M. majus</i>	CBS 741.79	Germany	KP858937	KP859001	KP859064	KP859110
	10099	France	–	JX280597	JX280563	JX280560
	10098	France	–	–	JX280564	JX280561
	99027	Canada	–	JX280583	JX280566	–
	200107	Norway	–	KT736191	KT736253	KT736287
<i>M. miscanthi</i>	SAUCC211092 ^T	China	OM957532	OM956214	OM981141	OM981148
	SAUCC211093	China	OM957533	OM956215	OM981142	OM981149
	SAUCC211094	China	OM957534	OM956216	OM981143	OM981150
<i>M. musae</i>	CBS 143500 ^T	Malaysia	MH107942	MH107895	MH108041	MH108003
	CBS 143499	Malaysia	MH107941	MH107894	MH108040	–

Species	Voucher	Country	GenBank Accession Number			
			LSU	ITS	tub2	rpb2
<i>M. musae</i>	CBS 111018	Costa Rica	–	AY293061	–	–
	CPC 11240	Mauritius	MH107944	MH107897	MH108043	–
	CPC 16258	Mexico	MH107945	MH107898	MH108044	–
	CPC 11234	Mauritius	MH107943	MH107896	MH108042	–
	CPC 32681	Malaysia	MH107946	MH107899	–	–
<i>M. neoqueenslandicum</i>	CBS445.95	The Netherlands	KP858933	KP858997	KP859060	KP859106
	CBS108926 ^T	New Zealand	KP858938	KP859002	KP859065	KP859111
<i>M. nivale</i>	CBS 116205 ^T	UK	KP858944	KP859008	KP859071	KP859117
	200114	Norway	–	KT736185	–	KT736279
	200119	Norway	–	KT736199	KT736240	KT736263
	200120	Norway	–	KT736210	KT736221	KT736273
	200566	Norway	–	KT736220	KT736224	–
	201050	Norway	–	KT736217	KT736236	KT736257
<i>M. novae-zelandiae</i>	CBS 143847	New Zealand	–	LT990655	LT990608	LT990641
	CPC 29693	New Zealand	–	LT990656	LT990609	LT990642
<i>M. paspali</i>	CBS 138620 ^T	China	–	KJ569509	KJ569514	–
	CBS 138621	China	–	KJ569510	KJ569515	–
	CBS 138622	China	–	KJ569511	KJ569516	–
<i>M. phragmitis</i>	CBS 285.71 ^T	Poland	KP858949	KP859013	KP859077	KP859122
	CBS 423.78	Germany	KP858948	KP859012	KP859076	KP859121
<i>M. poae</i>	CGMCC3.19170 ^T	China	–	MH740898	MH740914	MH740906
	LC12115	China	–	MH740901	MH740917	MH740909
	LC12116	China	–	MH740902	MH740918	MH740910
	LC12117	China	–	MH740903	MH740919	MH740911
	LC12118	China	–	MH740897	MH740913	MH740905
	LC12119	China	–	MH740899	MH740915	MH740907
	LC12120	China	–	MH740904	MH740920	MH740912
	LC12121	China	–	MH740900	MH740916	MH740908
<i>M. ratticaudae</i>	BRIP 68298 ^T	Australia	MW481666	MW481661	–	MW626890
<i>M. rhopalostylidis</i>	CBS 145125 ^T	New Zealand	MK442532	MK442592	MK442735	MK442667
<i>M. salmonicolor</i>	NC14-294	South Korea	MK836108	MK836110	–	–
<i>M. seminicola</i>	CBS 122706	Switzerland	KP858943	KP859007	KP859070	KP859116
	CBS 122707	Switzerland	KP858947	KP859011	KP859081	KP859120
	CBS 139951 ^T	Switzerland	KP858974	KP859038	KP859101	KP859147
	KAS1516	Canada	KP858961	KP859025	KP859088	KP859134
	KAS3574	Switzerland	KP858973	KP859037	KP859100	KP859146
	KAS3158	Canada	KP858970	KP859034	KP859097	KP859143
	KAS1527	Canada	KP858966	KP859030	KP859093	KP859139
	KAS1473	Canada	KP858955	KP859019	KP859082	KP859128
<i>M. shilinense</i>	CGMCC 3.23531 ^T	China	OP104022	OP103972	OP242834	–
<i>M. sinense</i>	SAUCC211097 ^T	China	OM959225	OM956289	OM981144	OM981151
	SAUCC211098	China	OM959226	OM956290	OM981145	OM981152
<i>M. sorghi</i>	CBS 691.96	Cuba	KP858936	KP859000	KP859063	KP859109
<i>Microdochium</i> sp.	SAUCC1017	China	MT199879	MT199885	MT435654	–
<i>M. tainanense</i>	CBS 269.76 ^T	Taiwan	KP858945	KP859009	KP859072	KP859118
	CBS 270.76	Taiwan	KP858931	KP858995	KP859058	KP859104

Species	Voucher	Country	GenBank Accession Number			
			LSU	ITS	tub2	rpb2
<i>M. trichocladiopsis</i>	CBS 623.77 ^T	Unknown country	KP858934	KP858998	KP859061	KP859107
<i>M. triticolae</i>	RR 241	UK	–	AJ748691	–	–
<i>M. chrysopogonis</i>	GDMCC 3.683	China	MT988024	MT988022	MW002441	MW002444
	LNU-196	China	MT988023	MT988020	MW002442	MW002445
	LNU-197	China	MT988025	MT988021	MW002443	MW002446
<i>M. yunnanense</i>	SAUCC1018	China	MT199880	MT199886	MT435655	–
	SAUCC1015	China	MT199877	MT199883	MT435652	MT510549
	SAUCC1012	China	MT199876	MT199882	–	MT510548
	SAUCC1011 ^T	China	MT199875	MT199881	MT435650	MT510547
<i>Thamnomycetes dendroidea</i>	CBS 123578	France	KY610467	FN428831	KY624313	KY624232

Note: “T” denotes ex-type strain. Newly-generated sequences are indicated in bold. “–” means no data available in GenBank.

Phylogenetic analyses

The sequences of the strains from *C. zizanioides* and those of *Microdochium* species, as well as the outgroup *Idriella lunata* obtained from NCBI GenBank, were aligned with MAFFT version 7 using the default settings. Manual adjustments were made to optimise the alignment in MEGA version 7.0 (Kato and Standley 2013; Kumar et al. 2016). To elucidate the taxonomic phylogenetic relationships, single and concatenated ITS, LSU, *rpb2* and *tub2* sequence alignments were subjected to analysis by applying Bayesian Inference (BI) using MrBayes version 3.2.5 and Maximum Likelihood (ML) using RAxML on the CIPRES portal (www.phylo.org) (Swoford 2002; Crous et al. 2006; Ronquist et al. 2012). For BI analysis, the best evolutionary model was determined through the utilisation of MrModelTest version 2.2 (Nylander 2004). Subsequently, in MrBayes v. 3.2.5, the Markov Chain Monte Carlo 180 (MCMC) algorithm was used to generate phylogenetic trees. The first 25% of saved trees were discarded as the burn-in phase. Posterior probabilities (PPs) were determined from the remaining trees prior to calculation of the 50% majority rule consensus trees; PP values exceeding 0.90 were considered significant. The ML analyses were performed by using RAxML-HPC BlackBox version 8.2.6, based on 1000 bootstrap replicates. A general time reversible (GTR) model was applied with gamma-distributed rate variation. Bootstrap values (BSs) equal to or higher than 70% were regarded as significant. The phylogenetic tree was viewed in Fig-Tree version 1.4.4 (Rambaut 2018) and edited by Adobe Illustrator CC2018.

Pathogenicity test

Three isolates of *M. chrysopogonis* (GDMCC 3.683, LNU-196 and LNU-197) were used to conduct the pathogenicity test. *C. zizanioides* plants were cultivated within a greenhouse, utilising plastic pots containing field-collected soil from the location where the plants had been established. The isolates were cultured on PDA for 2 weeks at 30 °C in the dark to collect conidia. For the detached leaf assay, 1-cm wide leaves were harvested from 2-month-old plants cultivated in a greenhouse, washed under running tap water, surface disinfected with 70% ethanol for 1 min, rinsed with sterile water for 30 seconds and finally air-dried on sterilised filter paper. The conidial suspension was adjusted to a concentration of 1 × 10⁶ conidia/ml in sterile distilled water. An equivalent volume

of sterile distilled water was used as a control. Leaf blades were then wounded with a sterilised pin and each leaf was sprayed with 2 ml of conidial suspension. All inoculated and control leaves were placed in a moist chamber at 25 °C with 100% relative humidity (RH) under cool fluorescent light with a 12-h photoperiod. After seven days, the disease incidence was assessed and calculated as the percentage of leaves with leaf tar spot symptoms. Each treatment consisted of five replicates and the experiment was conducted three times.

For the attached leaf assay, the leaf blades of healthy leaves were also pin-pricked and the conidial suspension was adjusted to a concentration of 2×10^6 conidia/ml in sterile distilled water. An equivalent volume of sterile distilled water was used as a control. In each treatment, five plants were included, with each plant being sprayed with approximately 20 ml of inoculum. All sprayed and control plants were incubated in a plastic container in a greenhouse at 25 ± 2 °C under cool fluorescent light with a 12-h photoperiod. For the first 3 days, the plastic container was covered with transparent polyethylene bags to maintain a high humidity. The disease incidence was assessed 10 days post inoculation and calculated as the percentage of plants displaying tar spot symptoms. Each treatment had three replicates and the pathogenicity test was repeated twice.

To fulfil Koch's postulates, symptomatic leaf tissues were subjected to surface sterilisation as described above. Subsequently, these tissues were plated on to PDA medium to enable the re-isolation of the fungi. These isolates were identified, based on comparison of the cultures with those of the original strains. Furthermore, the identifications were confirmed by sequencing of the isolates.

Effect of temperature on mycelial growth rate

Mycelial growth rates of *M. chrysopogonis* isolates were assessed across various temperatures. Mycelial plugs with a diameter of 5 mm were excised using a sterile hole puncher from the periphery of 10-day-old PDA cultures. Subsequently, they were translocated to the central area of 90 mm PDA Petri dishes. The cultures were subjected to incubation across a temperature range of 5, 10, 15, 20, 25, 30, 35, 40 and 45 °C. Four replicate plates per isolate were prepared for each temperature. The plates were enveloped using Parafilm (Bemis Company, Neenah, WI, U.S.A.) and then positioned within plastic containers prior to their placement in incubators. The colony diameter was measured along two mutually perpendicular axes and the mean of these two measurements was documented as the radial colony diameter. Following a 10-day duration, mycelial growth rates were determined, based on colony diameter and subsequently quantified in millimetres per day. Each treatment was replicated four times.

Fungicide sensitivity

To determine possible control measures for this pathogen in the field, six groups including nine fungicides were tested for their ability to inhibit the growth of *M. chrysopogonis* in vitro. Fungicide sensitivity assays were conducted, based on methods developed by Yin et al. (2019). The commercial formulations of fungicides were serially diluted using sterilised distilled water. These diluted solutions were added to autoclaved PDA medium that had been cooled to 55 °C to obtain the desired concentrations in micrograms per millilitre (Table 2). Isolates were cul-

tured on PDA plates at 30 °C for 10 days in darkness to supply inoculum. Mycelial discs (5 mm in diameter) from the periphery of colonies actively growing on PDA were positioned at the centre of the fungicide-amended plates and unamended (control) plates. The plates were then incubated at a temperature of 30 °C in darkness for a duration of 2 weeks. Subsequently, the diameter of each colony was measured along two perpendicular axes and the mean diameter was recalibrated by deducting the diameter of the original plug utilised for inoculation. The effective concentration for 50% mycelial growth inhibition (EC_{50}) was estimated by performing a regression analysis of the percentage of mycelial growth inhibition against the \log_{10} of fungicide concentrations. Each treatment was replicated four times.

Statistical analysis

The dataset was tested for variance homogeneity using the Levene test. If the variances were equal, an analysis of variance (ANOVA) followed by a least significant difference (LSD) test was conducted. In cases where the variances were unequal, the Dunnett T3 test was applied. All statistical analyses were carried out using IBM SPSS version 20.0 (SPSS Inc., Chicago, IL, U.S.A.). The significance threshold for detecting treatment disparities was set at $P < 0.05$.

Results

Disease symptoms and isolation of the pathogen

From 2019 to 2022, a previously unknown disease of vetiver grass occurred during late spring and early autumn at the Grass Research Station of Lingnan Normal University (LNU) in Guangdong Province, China. Symptoms consistently appeared on 85% of *C. zizanioides* grown under field conditions. The initial symptoms appeared as small and scattered punctate spots (< 1 cm) embedded within the leaf tissue. Gradually, these spots clustered on leaf surfaces. Subsequently, brown, elliptical, fish-eye necrotic haloes emerged, encircling the lesion spots and aligning parallel to the leaf veins (Fig. 1A–C). As these necrotic haloes coalesced, the leaf underwent chlorosis and wilting, eventually leading to blighting of the entire plant. Ascomata were visible on the diseased leaf surfaces.

Table 2. List of the fungicides used in this study.

Active ingredient	Chemical family	Trade name	FRAC code	Concentration (µg/ml)
Pyrimethanil	anilino-pyrimidines	Syngenta	9	0, 0.01, 0.1, 1, 10, 100
Difenoconazole	triazoles	Syngenta	3	0, 0.01, 0.1, 1, 10, 100
Fludioxonil	phenylpyrroles	Syngenta	12	0, 1, 10, 30, 100, 300
Iprodione	dicarboximides	Syngenta	2	0, 1, 10, 30, 100, 300
Flusilazole	triazoles	Syngenta	3	0, 0.001, 0.01, 0.1, 1, 10
Propiconazole	triazoles	BASF	3	0, 0.0016, 0.008, 0.04, 0.2, 1
Carbendazim	benzimidazoles	Syngenta	1	0, 0.0016, 0.008, 0.04, 0.2, 1
Metalaxyl	acylalanines	BASF	4	0, 10, 30, 100, 300, 1000
Tebuconazole	triazoles	Bayer	3	0, 0.0016, 0.008, 0.04, 0.2, 1

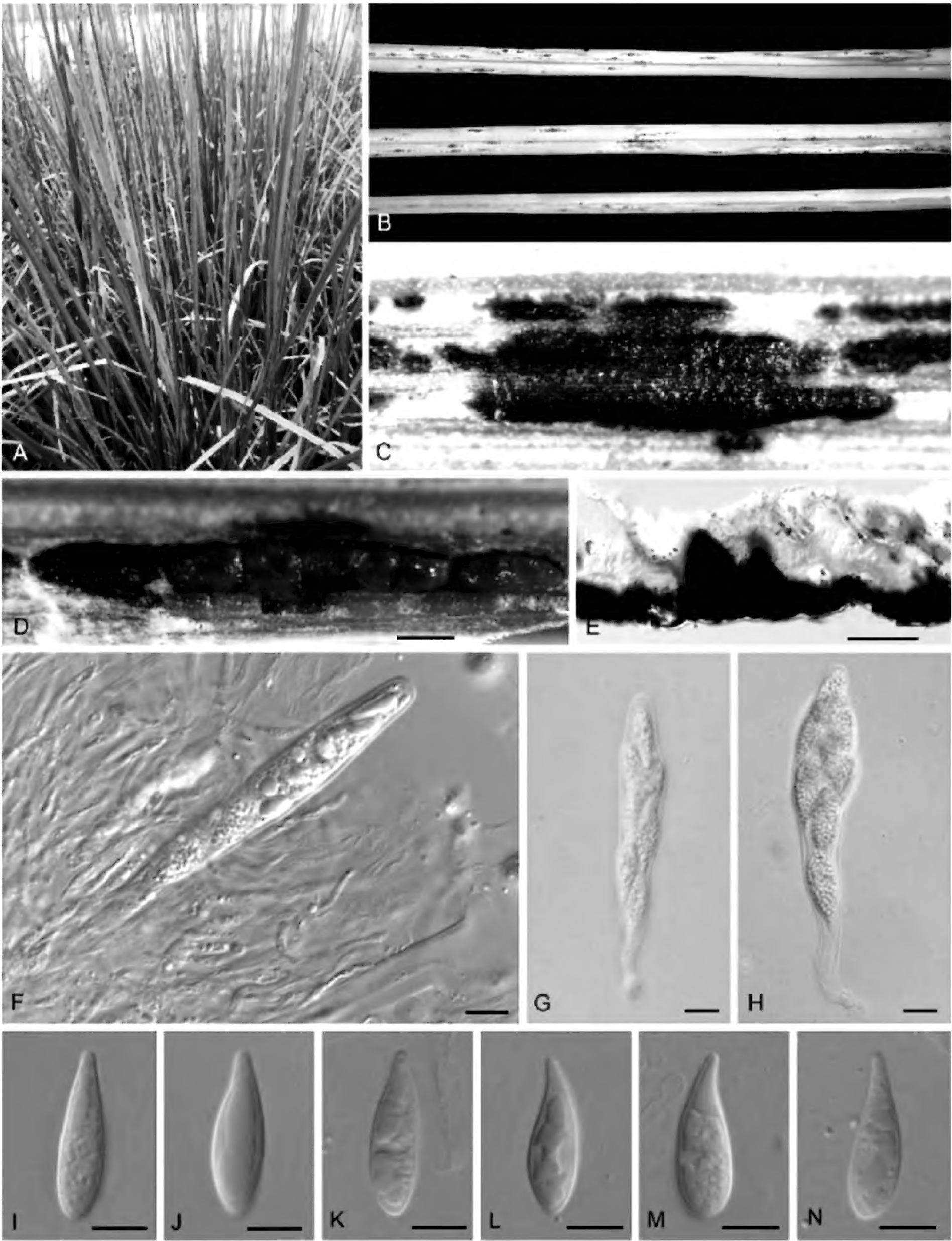


Figure 1. Disease symptoms and morphological characters of *Microdochium chrysopogonis* on infected leaf tissue (CAF 800053) **A–C** tar spot symptoms of *Chrysopogon zizanioides* from natural infection in the field **D** appearance of immersed ascomata on infected leaves **E** ascomata in longitudinal section **F–H** asci **I–N** ascospores. Scale bars: 100 μ m (**D, E**); 10 μ m (**F–H**).

A total of 67 isolates were obtained on PDA. As the colony morphology of the isolates was consistent, three representative isolates (GDMCC 3.683, LNU-196 and LNU-197), one from each field, were selected for further studies.

Phylogeny

Based on a Megablast search on NCBI's GenBank nucleotide database, the closest hits for the ITS sequence of strain GDMCC 3.683 were *M. dawsoniorum* sequences with 98% identity (538/551, MK966337; 532/543, MN492650) and a *Microdochium* sp. sequence with 97% identity (545/562, FJ536210). The closest hits for LSU sequence of this strain were *M. dawsoniorum* sequences with 99% identity (868/871, OM333563; 864/867, ON394569) and a *M. yunnanense* sequence with 99% identity (875/882, MT199880). The closest hits for its *rpb2* sequence were *M. tainanense* sequences with 85% identity (711/841, KP859118 and KP859104) and a *M. neoqueenslandicum* sequence with 83% identity (698/842, KP859106). The closest hits for *tub2* sequence were a *M. tainanense* sequence with 95% identity (661/697, KP859058), a *M. neoqueenslandicum* sequence with 95% identity (665/703, KP859060) and a *M. colombiense* sequence with 95% identity (658/695, KP859062). Therefore, molecular analyses with all available *Microdochium* species were performed. The alignment of each single locus and concatenated sequence dataset of ITS, LSU, *rpb2* and *tub2* were used to confirm species resolution in *Microdochium*.

There were in total 99 aligned sequences, including the outgroup, *Thamnomycetes dendroidea*. A total of 3,033 characters (547 bp from the ITS, 843 bp from LSU, 848 bp from *tub2* and 795 bp from *rpb2*) were included in the phylogenetic analyses. RAxML analysis of the combined dataset yielded a best scoring tree with a final ML optimisation likelihood value of -21, 329.537402 (ln). The matrix had 1,096 distinct alignment patterns with 27.41% undetermined characters or gaps. The tree length was 3.410120. Estimated base frequencies were: A = 0.234382, C = 0.267827, G = 0.258835, T = 0.238956; substitution rates were AC = 1.101009, AG = 4.781387, AT = 1.240884, CG = 0.955029, CT = 6.933148 and GT = 1.000000; gamma distribution shape parameter α = 0.152657. Based on the results of MrModelTest, the SYM + I + gamma for ITS, GTR + I + gamma for LSU and *rpb2* and HKY + I + gamma model for *tub2* were selected as the best fit models for Bayesian analyses. A total of 47,402 trees were generated by BI, amongst which 11,851 trees were discarded as the burn-in phase and the remaining 35,551 trees were used to calculate the posterior probabilities (PPs). The BI consensus tree confirmed the tree topology obtained with ML. The well-supported clade (1/100) formed by the three strains from *C. zizanioides* clustered with high support (1/100) with *M. dawsoniorum* (0.92/97), which was sister to one single-strain clade representing *M. ratticaude*. This clade clustered with high support (0.92/93) with the clade formed by *M. albes-cens*, *M. seminicola*, *M. graminearum*, *M. shilinense*, *M. insulare*, *M. paspali*, *M. citrinidiscum*, *M. sorghi*, *M. tainanense* and *M. trichocladiopsis* strains. The *M. neoqueenslandicum* clade (1/100) was basal to this clade (Fig. 2). Single gene-based phylogenies are presented in the Suppl. material 1. Nonetheless, these individual gene trees did not yield a conclusive taxonomic classification for the new species, in contrast to the comprehensive resolution achieved through the concatenated sequence analysis.

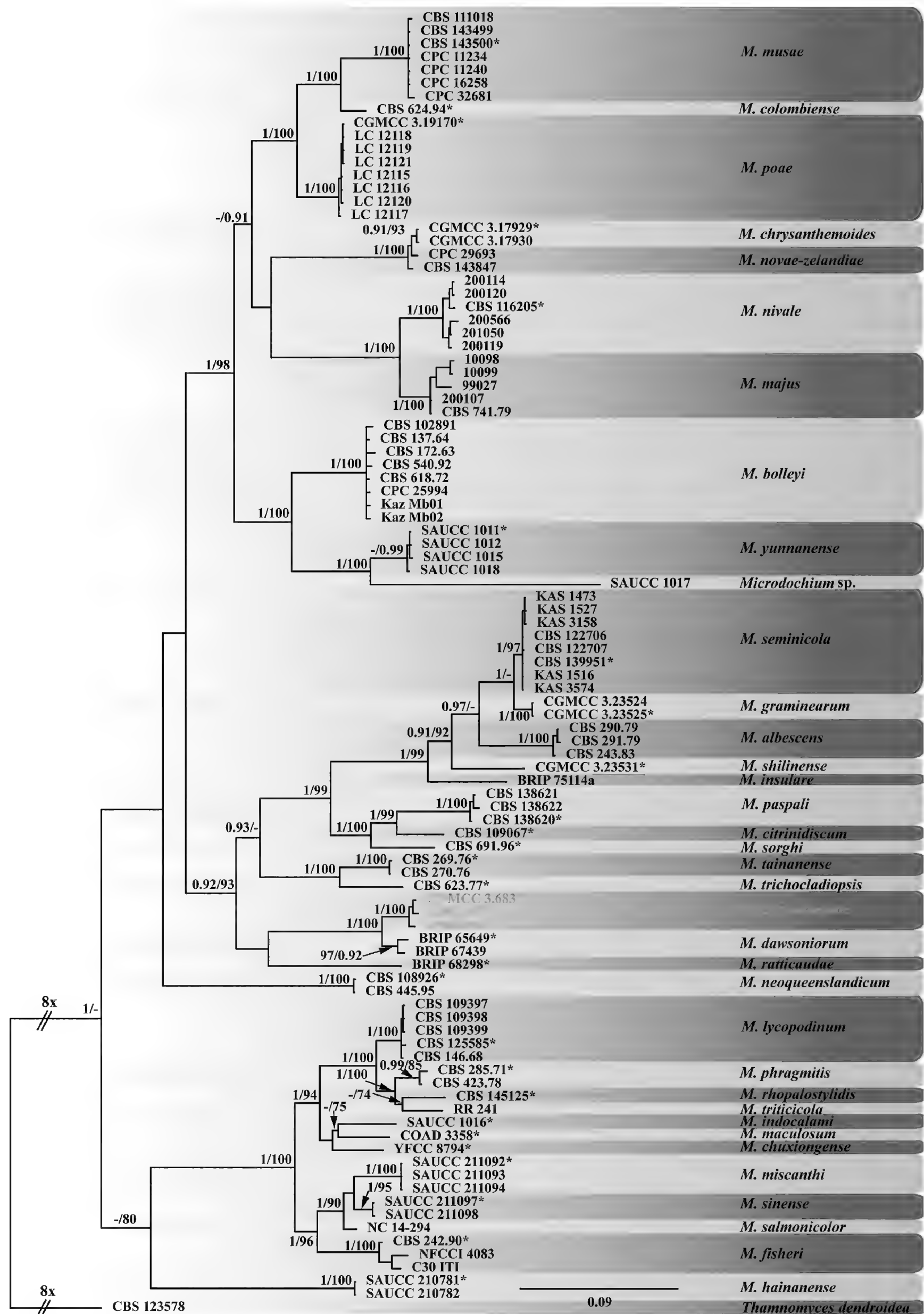


Figure 2. Phylogenetic tree inferred from a Maximum Likelihood analysis, based on a combined alignment of ITS, LSU, *tub2* and *rpb2* sequences from 99 isolates of *Microdochium* sp. Bootstrap support values obtained with ML above 70% and Bayesian (BI) posterior probability values above 0.90 are shown at the nodes (BI/ML). The tree was rooted to *Thamnomycetes dendroidea* CBS 123578. Numbers of ex-type strains are emphasised with an asterisk and species are delimited with shaded blocks. Isolates of *M. chrysopogonis* are indicated with lighter text.

Taxonomy

Based on multilocus phylogenetic analyses, the three strains isolated from *C. zizanioides* represent a previously unknown species within the genus *Microdochium* that is closely related to *M. dawsoniorum* and *M. ratticaudae*. Morphological data placed the new species in the genus *Microdochium*. This species is characterised below.

Microdochium chrysopogonis W. Zhang & X. Lu, sp. nov.

MycoBank No: 845624

Figs 1, 3

Etymology. Name refers to *Chrysopogon*, the host genus from which this fungus was collected.

Description. **Sexual morph** on infected leaf tissue of the host plant (CAF 800054). **Ascomata** perithecial, 300–350 µm diam., solitary or in groups, immersed, pale brown to black, subglobose to oval, uniloculate, non-ostiolate. **Paraphyses** filiform, hyaline, straight or curved, apically free. **Asci** 50–60 × 10–18, $\bar{x} = 55 \times 13$ µm ($n = 50$), hyaline, fasciculate, unitunicate, oblong to narrowly clavate, fusiform, 8 biseriate spores with a short stipe. **Ascospores** clavate, hyaline, guttulate, 20–22 × 8–11.5, $\bar{x} = 21 \times 9$ µm ($n = 50$), aseptate, smooth. **Sporodochia** salmon-pink, slimy. **Conidiophores** reduced to conidiogenous cells. **Conidiogenous cells** with percurrent proliferation, hyaline, smooth, aseptate, ampulliform or obpyriform, 10–23 × 8–11.5, $\bar{x} = 17 \times 9.5$ µm ($n = 50$). **Conidia** fusiform, lunate, curved, solitary, guttulate, variable in length, 0–1-septate, 18–72 × 2–3.5, $\bar{x} = 38.5 \times 3$ µm ($n = 50$), apex rounded, base usually flattened. **Chlamydospores** not observed. Vegetative hyphae on PDA (GDMCC 3.683) superficial and immersed, septate, branched, hyaline, smooth, 1–5.5 µm wide.

Culture characteristics. Colonies on PDA reaching 4.0–4.5 cm within seven days in the dark at 30 °C, flat, white cottony aerial mycelium, dense, saffron rounded sporodochia produced after 3 weeks; reverse saffron. On MEA, sparse white cottony aerial mycelium, orange rounded sporodochia produced; reverse salmon-pink. On OA, periphery with white scarce cottony aerial mycelium, concentric rings of orange rounded sporodochia produced; reverse orange.

Type. China, Guangdong Province, Zhanjiang City, field of the Grass Research Station of Lingnan Normal University (LNU), from a leaf of vetiver grass (*Chrysopogon zizanioides*) with leaf tar spot disease, September 2019, W. Zhang & X. Lu, holotype CAF 800054, ex-type living strain GDMCC 3.683.

Additional materials examined. China, Guangdong Province, Zhanjiang City, field of the Grass Research Station of Lingnan Normal University (LNU), from a leaf of vetiver grass (*C. zizanioides*) with leaf tar spot disease, September 2019, W. Zhang & X. Lu, strain LNU-196; China, Guangdong Province, Zhanjiang City, field of the Grass Research Station of Lingnan Normal University (LNU), from a leaf of vetiver grass (*C. zizanioides*) with leaf tar spot disease, September 2019, W. Zhang & X. Lu, strain LNU-197.

Notes. A multilocus phylogenetic analysis of the ITS, LSU, *tub2* and *rpb2* loci placed three strains of *M. chrysopogonis* in a distinct and monophyletic clade (1/100) sister to *M. dawsoniorum* and *M. ratticaudae*. Notably, *M. chrysopogonis*

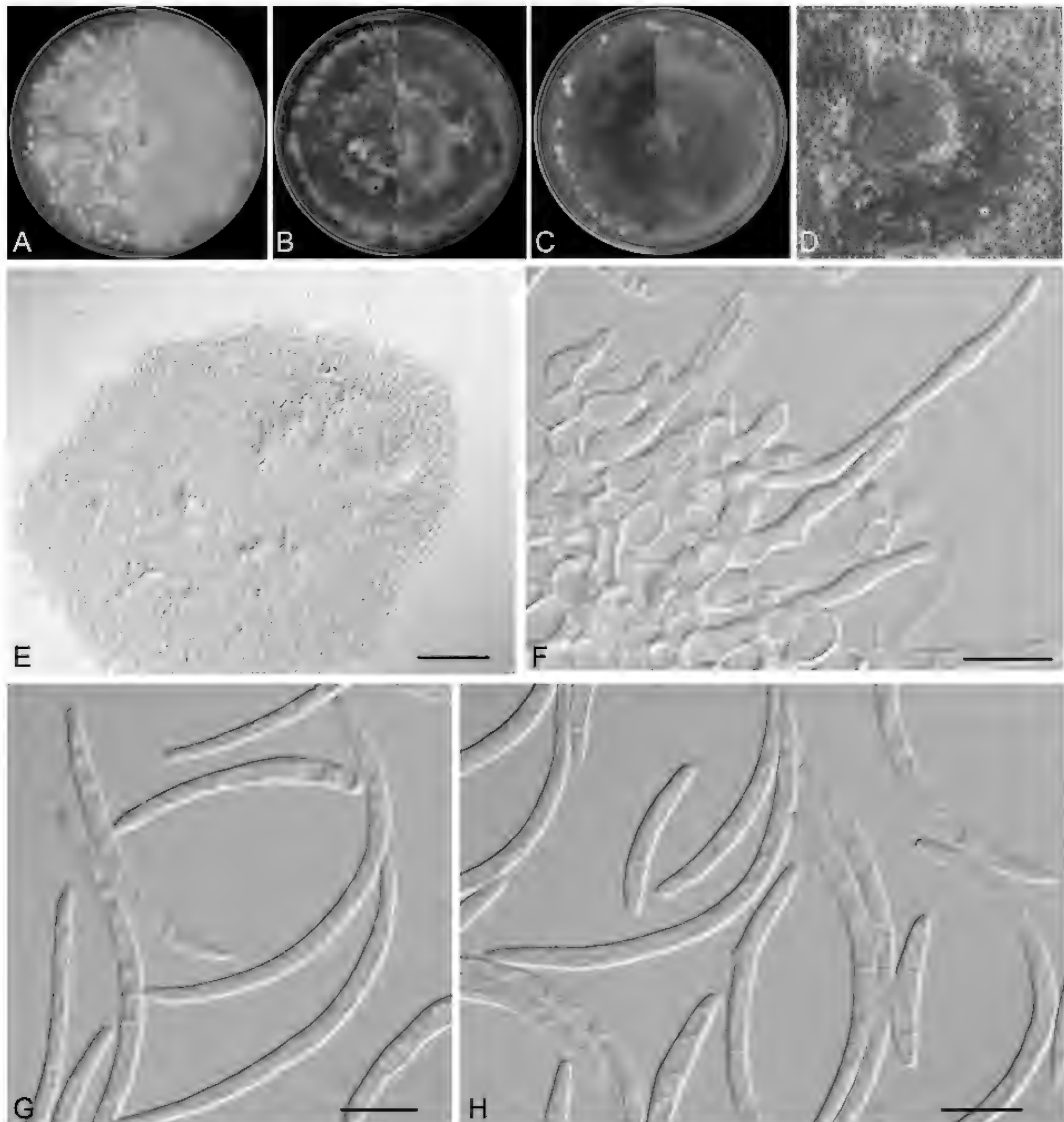


Figure 3. *Microdochium chrysopogonis* (from ex-type: GDMCC 3.683) **A** colonies after 7 days on PDA **B** colonies after 7 days on MEA **C** colonies after 7 days on OA **D** colony overview of the sporodochia on PDA in culture after incubation for three weeks **E** aggregated conidiophores **F** conidiophores with conidiogenous cells **G, H** conidia. Scale bars: 20 µm (**B**).

has longer conidia ($18\text{--}72 \times 2\text{--}3.5 \mu\text{m}$) than *M. ratticaudae* ($7\text{--}11 \times 1.5\text{--}2.5 \mu\text{m}$) and wider conidia than *M. dawsoniorum* ($25\text{--}75 \times 1\text{--}2 \mu\text{m}$). Furthermore, the conidia of *M. chrysopogonis* are guttulate and 0–1-septate, while those of *M. dawsoniorum* are 0–3-septate and those of *M. ratticaudae* are aseptate. The conidiogenous cells of *M. chrysopogonis* appear as percurrent, ampulliform or obpyriform, whereas those of *M. ratticaudae* are indistinct from the hyphae and those of *M. dawsoniorum* are cylindrical to irregular and flexuous. Additionally, the conidiogenous cells of *M. chrysopogonis* ($10\text{--}23 \times 8\text{--}11.5 \mu\text{m}$) are wider than those of *M. ratticaudae* ($20\text{--}30 \times 1\text{--}2 \mu\text{m}$) (Table 3). Differences are also evident in the sexual morph of these three species. In particular, the sexual

morph is not observed in *M. dawsoniorum*. Ascomata size varies, with that of *M. ratticaudae* (100–160 μm) being smaller than that of *M. chrysopogonis* (300–350 μm). Ascospores of *M. ratticaudae* (14–24 \times 4–7 μm) are fusoid to navicular, while those of *M. chrysopogonis* are clavate, guttulate and wider (20–22 \times 8–11.5 μm). In addition, *M. ratticaudae* features abundant, pale to olivaceous brown, subglobose or cylindrical chlamydospores, while these are not observed in *M. chrysopogonis* (Crous et al. 2020, 2021a; Table 3). Consequently, based on both morphological characteristics and phylogenetic analyses, all three isolates of *M. chrysopogonis* were proposed as a new species.

Pathogenicity test

The symptoms observed on leaves of *C. zizanioides* after inoculation with the representative isolate GDMCC 3.683 were similar to those observed in the field. No symptoms were observed on the leaves of the negative controls (Fig. 4). The average disease incidence of detached leaves that were wounded and sprayed with the isolates GDMCC 3.683, LNU-196 and LNU-197 was 93.3%, 80.0% and 93.3%, respectively. The average disease incidence of whole plants after spraying with the same isolates was 76.7%, 73.3% and 73.3%, respectively (Fig. 5). Koch's postulates were fulfilled by successful re-isolation of the fungal strains from all leaf spot tissues inoculated with the three isolates. The morphology and DNA sequences of the isolates re-isolated from the inoculated tissues were consistent with those of the strains used for inoculations.

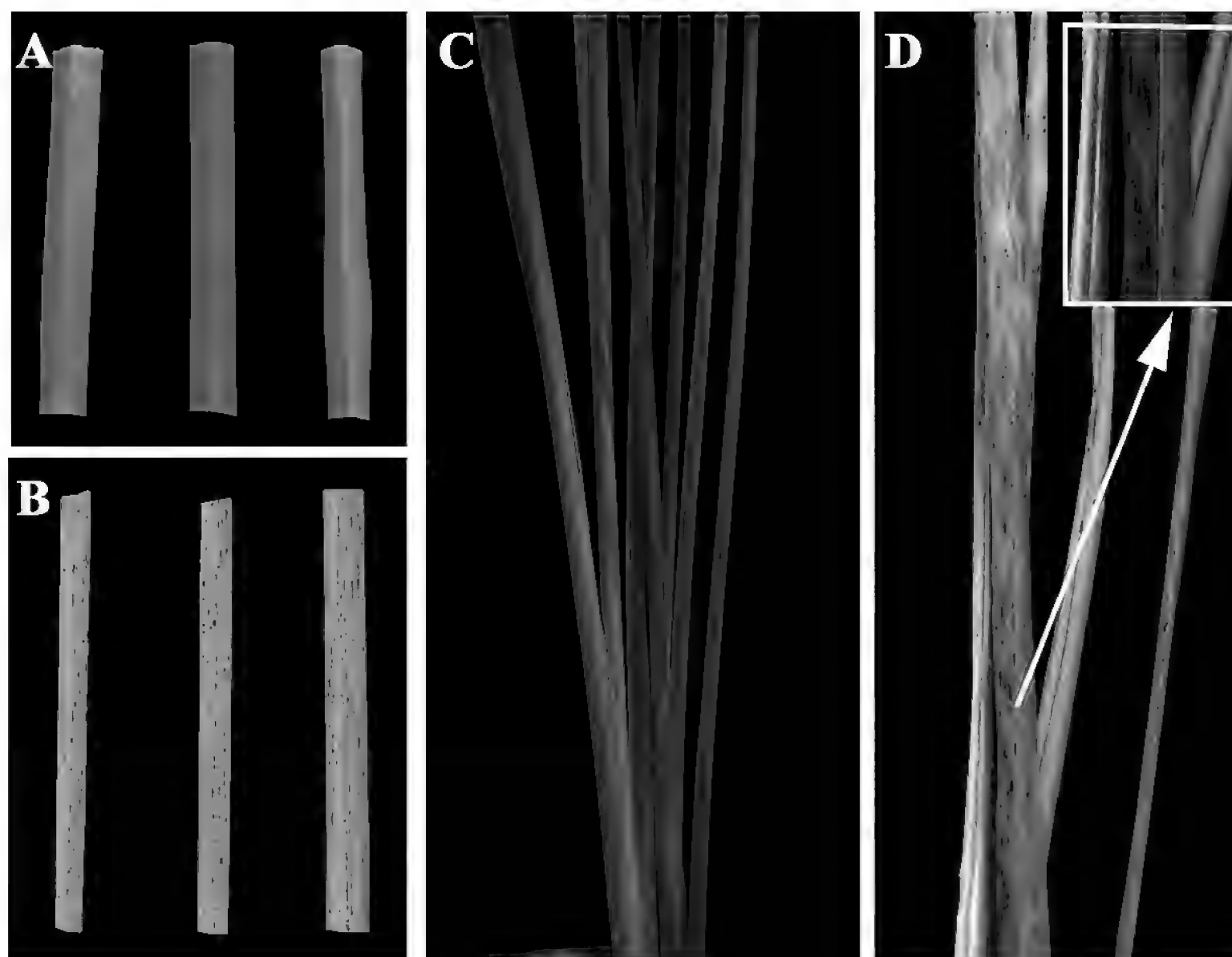


Figure 4. Tar spot symptoms of *Chrysopogon zizanioides* 7 days after spraying on detached leaves (**A**, **B**) and 10 days after spraying on leaves attached to whole plants (**C**, **D**) with *Microdochium chrysopogonis* isolate GDMCC 3.683 (**B**, **D**) and sterilised water (**A**, **C**).

Table 3. Morphological characters of *Microdochium chrysopogonis* and its related species.

Taxa		<i>M. albescens</i>	<i>M. citrinidis-cum</i>	<i>M. neoqueens-landicum</i>	<i>M. paspali</i>	<i>M. semini-cola</i>	<i>M. tricho-cladiopsis</i>	<i>M. tainan-ense</i>	<i>M. sorghi</i>	<i>M. dawsonio-rum</i>	<i>M. ratticaudae</i>	<i>M. gramin-earum</i>	<i>M. shilin-ense</i>	<i>M. chrys-opogonis</i>	
Asexual morph	Conidia	falcate, slightly to strongly curved, apex pointed	cylindrical, clavate, obovoid	lunate, allantoid, curved	falcate, apex pointed	cylindrical to fusiform, straight or curved	oblong, fusiform to obovoid, straight or curved	lunate	filiform, narrowly acicular fusiform, obclavate	flexuous to falcate, sometimes with a geniculation, acute at the tip, narrow at the base	fusoid, falcate, acute at the apex and narrowed at the base	n/a	n/a	fusiform, lunate, curved, guttulate	
		Size (µm)	11–16 × 3.5–4.5	7–31 × 2–3	4–9 × 1.5–3	7–20.5 × 2.5–4.5	19–54 × 3–4.5	6–18 × 2–3.5	10–15 × 2–3	20–90 × 1.5–4.5	25–75 × 1–2	n/a	n/a	18–72 × 2–3.5	
		Septa	0–1(–3)	0–3	0(–1)	0–3	0(–3)3(–5)	0(–1)	0–1	1–7(–10)	0–3	aseptate	n/a	0–1	
	Conidioge-nous cells	Shape	subcylindrical, doliform to obpyriform	denticulate, cylindrical	ampulliform, lageniform to subcylindrical	ampulliform, lageniform to cylindrical	ampulliform to lageniform	cylindrical to clavate, straight but often curved at the tip	sympodial, apical, cylindrical or ampulliform with conspicuous rhachides	sympodial, ovoid, ampulliform to obclavate	cylindrical to irregular, flexuous, narrowed towards the tip	indistinct from hyphae, terminal, solitary.	n/a	n/a	ampulliform or obpyriform
Sexual morph		Size (µm)	6–15 × 1.5–4	11–29 × 1.5–2	4.5–10 × 2–3.5	6.5–15.5 × 2.5–4	7–9.5 × 3–4	4–37 × 2–3	3–10 × 1–3	5–13 × 3–4	20–30 × 1–2	n/a	16.3–22.4 × 4.1–5.7	10–23 × 8–11.5	
	Chla-mydo-spores	Shape	n/a	n/a	n/a	n/a	present	n/a	n/a	n/a	n/a	n/a	n/a	n/a	
	Perithecia	Size (µm)	150–180 × 90–120	n/a	n/a	n/a	110–149	n/a	n/a	n/a	n/a	n/a	n/a	300–350	
	Asci	Size (µm)	40–85 × 8–12	n/a	n/a	n/a	41–66 × 7.6–11	n/a	n/a	n/a	n/a	50–75 × 10–14	55–77.5 × 9.5–15.	50–60 × 10–18	
References	Ascospores	Size (µm)	14–23 × 3.5–4.5	n/a	n/a	n/a	12–22 × 3–4.5	n/a	n/a	n/a	14–24 × 4–7	16.5–24 × 4–5.5	14–18 × 3–5.5	20–22 × 8–11.5	
		Septa	1–3(–5)	n/a	n/a	n/a	0–3	n/a	n/a	n/a	aseptate	0–3	0–3	aseptate	
			Hernández- Restrepo et al. (2016)	Hernández- Restrepo et al. (2016)	Hernández- Restrepo et al. (2016)	Zhang et al. (2015) (Continued on next page)	Hernández- Restrepo et al. (2016)	Hernández- Restrepo et al. (2016)	De Hoog & Hermanides- Nijhof (1977)	Braun (1995)	Crous et al. (2020) (Continued on next page)	Crous et al. (2021a)	Gao et al. (2022)	Gao et al. (2022)	This study

Note: “n/a” means not provided in the literature.

Effect of temperature on mycelial growth

The mycelial growth of *M. chrysopogonis* was significantly affected by temperature ($P < 0.01$). All three isolates of *M. chrysopogonis* grew between 10 and 40 °C, with maximum growth observed at 30 °C (Fig. 6). No isolates grew at 5 or 45 °C after 3 days. The highest average mycelial growth rate was observed at 30 °C (26.5 ± 2.0 mm/day), followed by 25 °C (20.1 ± 4.7 mm/day).

Fungicide sensitivity

The EC_{50} values of various fungicides were analysed for their effectiveness against *M. chrysopogonis* isolates. A total of 17 isolates of *M. chrysopogonis* were collected from diseased leaves spanning the period from 2019 to 2022.

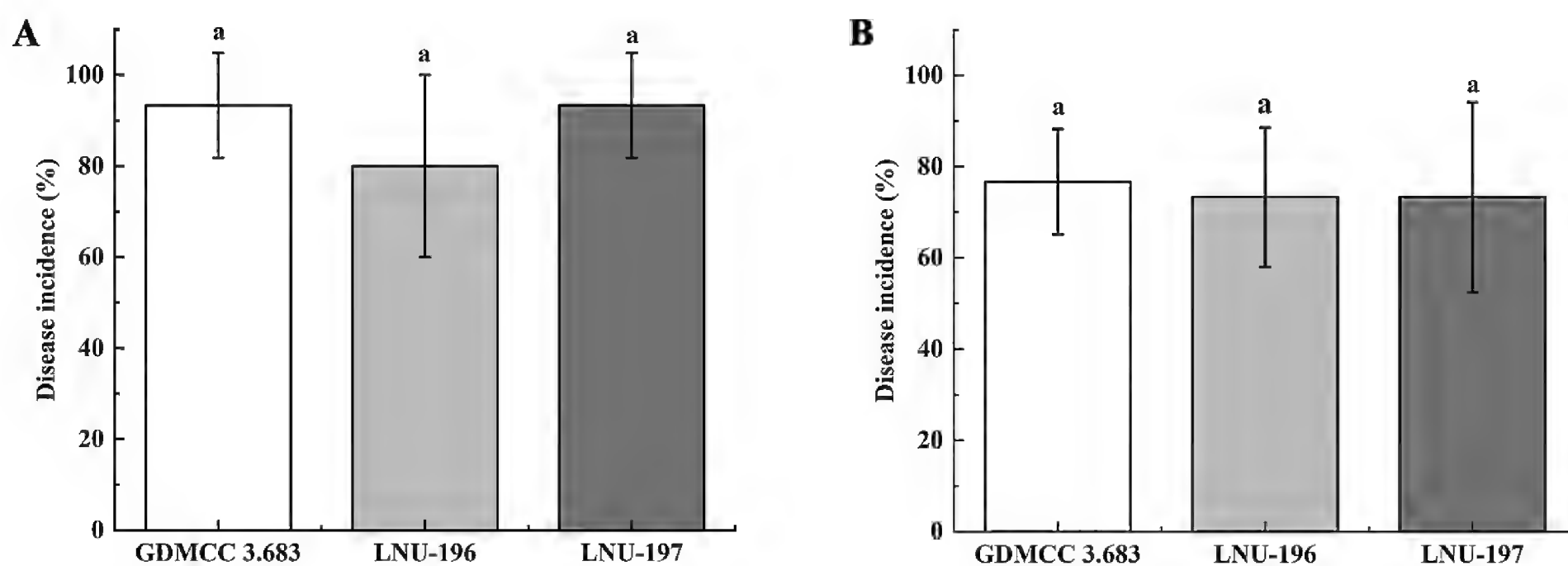


Figure 5. Disease incidence of tar spot symptoms on *Chrysopogon zizanioides* for leaves 7 days after spraying detached leaves (A) and for whole plants 10 days after spraying leaves attached to potted plants (B), respectively, with *Microdochium chrysopogonis* isolates GDMCC 3.683, LNU-196 and LNU-197. Values are shown as the means, with the error bars representing the standard error. For each pathogen, columns with the same letter indicate means that are not significantly different according to a least significant difference (LSD) test ($P < 0.05$).

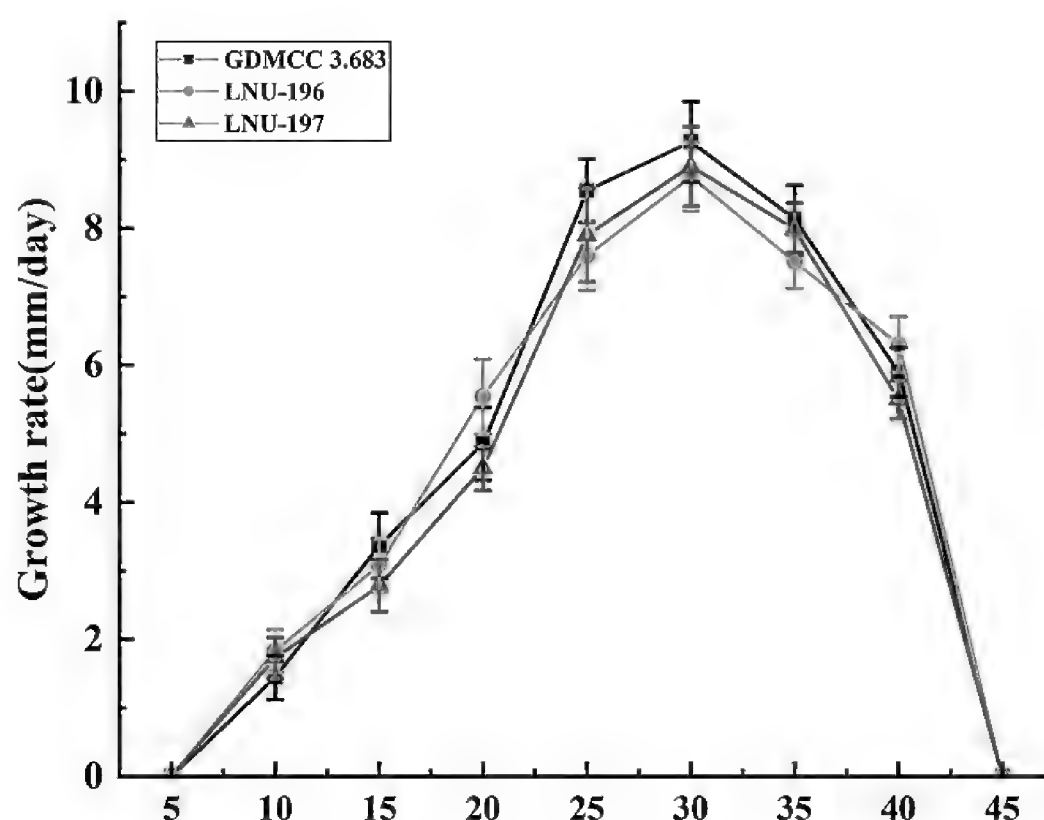


Figure 6. Colony growth rate of three isolates, GDMCC 3.683, LNU-196 and LNU-197, of *Microdochium chrysopogonis* from *Chrysopogon zizanioides* under different temperatures. Error bars represent the standard error.

The frequency distribution showed that difenoconazole, fludioxonil, flusilazole, carbendazim and iprodione exhibited distributions resembling normal curves, while pyrimethanil, propiconazole, metalaxyl and tebuconazole displayed unimodal curves (Fig. 7). EC₅₀ values for the inhibition of 17 *M. chrysopogonis* isolates, based on mycelial radial growth, varied across fungicide treatments ($P < 0.05$) (Table 4). Amongst the tested fungicides, flusilazole had the lowest EC₅₀ values, with a notably concentrated response range of 0.001 to 0.007 µg/ml and an average of 0.004 µg/ml. Tebuconazole closely followed with a slightly narrower range, exhibiting values ranging from 0.002 to 0.009 µg/ml and an average of 0.007 µg/ml. Propiconazole displayed EC₅₀ values spanning from 0.006 to 0.016 µg/ml, with an average of 0.011 µg/ml, while those of carbendazim ranged from 0.008 to 0.031 µg/ml, with an average of 0.024 µg/ml. In contrast, those of difenoconazole exhibited a broader range, varying from 0.013 to 0.127 µg/ml, with a mean value of 0.077 µg/ml, while those of pyrimethanil ranged from 0.054 to 0.605 µg/ml, with an average of 0.411 µg/ml. Those of ip-

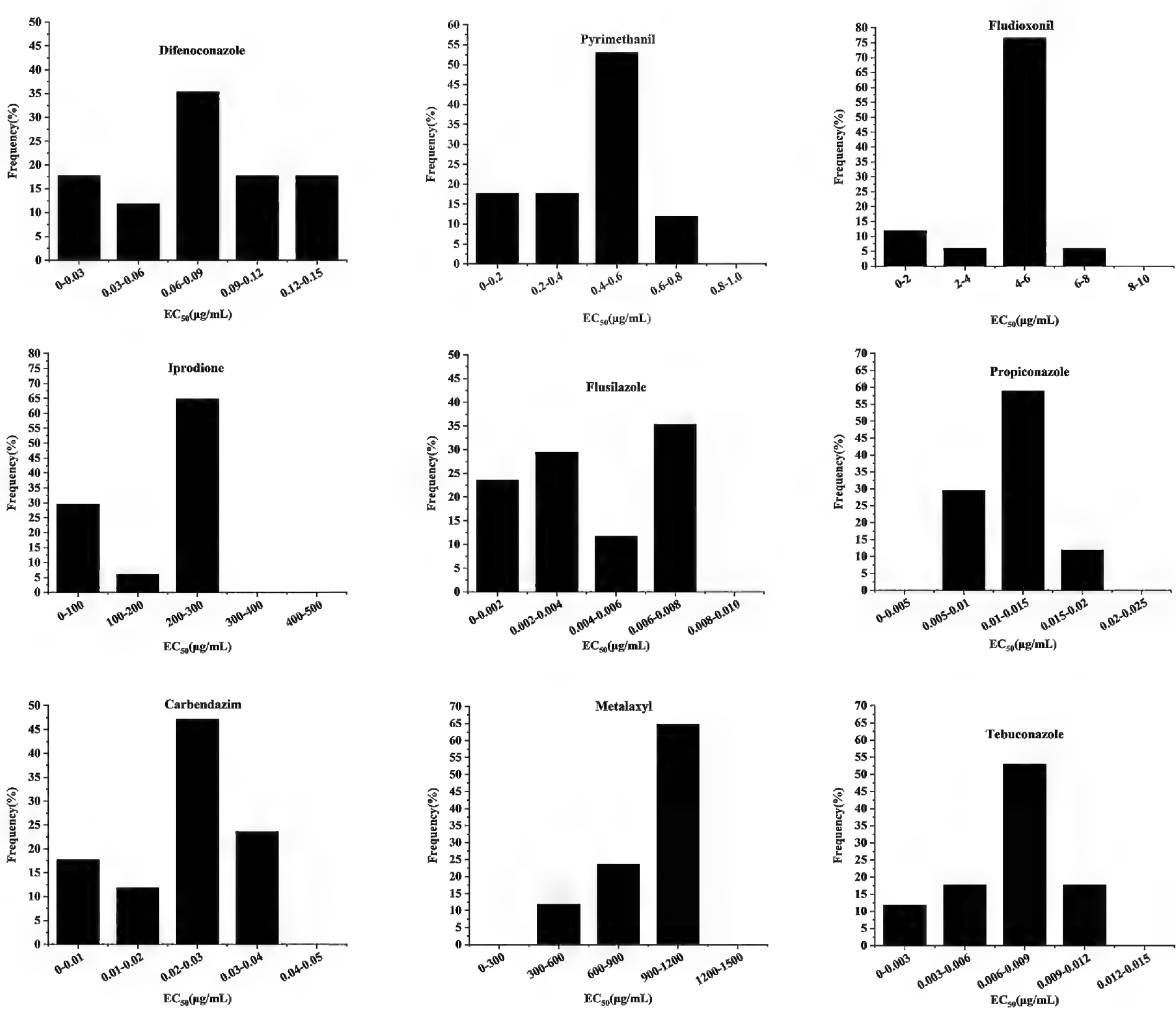


Figure 7. Frequency distribution of the 50% effective concentration (EC₅₀) values of six groups including nine fungicides for *Microdochium chrysopogonis* isolates, based on mycelial growth from 2019 to 2022.

Table 4. In vitro sensitivity ranges and mean 50% effective concentration (EC₅₀) values for the inhibition of *Microdochium chrysopogonis*.

Fungicide	EC ₅₀ (µg/ml)		
	Lowest	Highest	Mean ± SE
Difenoconazole	0.013	0.127	0.077 ± 0.039e
Pyrimethanil	0.054	0.605	0.411 ± 0.180d
Fludioxonil	0.014	6.128	4.525 ± 1.626c
Iprodione	15.018	260.335	193.031 ± 99.462b
Flusilazole	0.001	0.007	0.004 ± 0.003h
Propiconazole	0.006	0.016	0.011 ± 0.003g
Carbendazim	0.008	0.031	0.024 ± 0.009f
Metalaxyl	302.785	1056.896	892.677 ± 236.145a
Tebuconazole	0.002	0.009	0.007 ± 0.002h

Note: The letters indicate the comparison amongst the different fungicide treatments. Means followed by the same letter do not differ according to a post hoc Dunnett T3 test ($p < 0.05$).

rodione, on the other hand, spanned from 15.018 to 260.335 µg/ml, with an average of 193.031 µg/ml. Metalaxyl exhibited the highest EC₅₀ value, displaying the widest range amongst all fungicides, extending from 302.785 to 1056.896 µg/ml with an average of 892.677 µg/ml. Overall, these findings indicate varying degrees of sensitivity to different fungicides amongst *M. chrysopogonis* isolates. These variations in sensitivity could be essential considerations for designing effective fungicide application strategies against vetiver leaf tar spot disease.

The inhibition of mycelial growth revealed that all nine fungicides exhibited a reduction in fungal growth in vitro when compared to plates without amendments. The effectiveness of these fungicides in diminishing the mycelial growth of the isolates was contingent upon both the specific chemical compound and its concentration. Four DMI fungicides, namely, difenoconazole, propiconazole, flusilazole and tebuconazole and one MBC fungicide, carbendazim, displayed strong activity against *M. chrysopogonis* growth at concentrations below 1 µg/ml, specifically at concentrations of 1, 0.2, 0.1, 0.2 and 0.2 µg/ml, respectively (Fig. 8). However, *M. chrysopogonis* showed a tendency to exhibit better growth in the presence of pyrimethanil, fludioxonil, iprodione and metalaxyl, with mycelial growth being completely inhibited at concentrations exceeding 100 µg/ml (Fig. 8).

Discussion

In a survey of disease on *C. zizanioides* in Guangdong Province, China, from 2019 to 2022, tar spot was the predominant leaf spot disease. Isolation, morphological features, multilocus phylogenetic analysis and pathogenicity tests confirmed that a new *Microdochium* species, *M. chrysopogonis* was the causal agent. To effectively control the disease, the sensitivity of *M. chrysopogonis* to six groups of fungicides, including nine fungicides was determined. Results indicated that four DMI fungicides, namely difenoconazole, propiconazole, flusilazole and tebuconazole and one MBC fungicide, carbendazim, were highly effective against the new species.

The morphology of the new species is introduced along with its sexual and asexual morphological features, which are consistent with the following of *Microdochium*: pale brown to black, subglobose to oval, uniloculate, perithecial

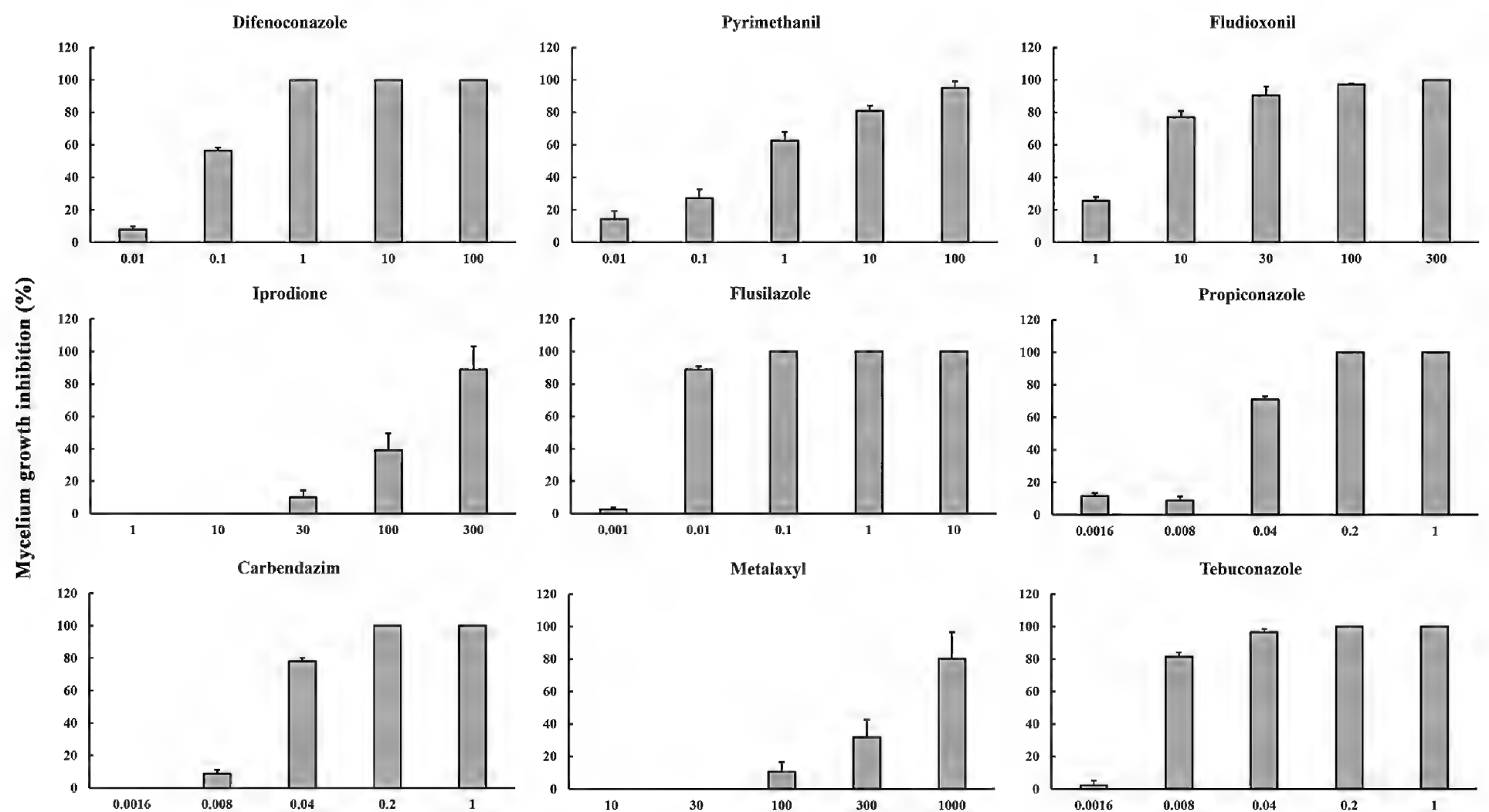


Figure 8. Effect of fungicides on the mycelial growth of *Microdochium chrysopogonis*. Values are shown as the means, with the error bars representing the standard error.

ascomata; hyaline, fasciculate, unitunicate, oblong to narrowly clavate, eight biserial spores with short stipe asci, from which hyaline, clavate, smooth ascospores arise. Conidiophores reduced to hyaline, smooth, aseptate, percurrent, ampulliform or obpyriform, conidiogenous cells, from which hyaline, 0–1-septate, fusiform, lunate conidia with the apex rounded and base flattened usually arise (Figs 1, 3) (Hernández-Restrepo et al. 2016). The concatenated ITS, LSU, *tub2* and *rpb2* sequences were able to identify species in *Microdochium* and proved to be suitable barcoding markers in the process of species resolution (Hernández-Restrepo et al. 2016). Phylogenetic analysis indicated that *M. chrysopogonis* formed a distinct well-supported clade (1/100) and was closely related to *M. dawsoniorum* and *M. ratticaudae* (Fig. 2). Nevertheless, the classification of the new species in the genus *Microdochium* is well supported by morphology, based on sexual and asexual morphs, which are different from those of *M. dawsoniorum* and *M. ratticaudae*.

Temperature is a major factor affecting plant disease epidemics. In recent years, tar spot disease of *C. zizanioides* has become increasingly prevalent in Guangdong Province, China, especially in hot and rainy summers. Thus, the effect of temperature on the growth rate of *M. chrysopogonis* in vitro was evaluated in this study. There were no significant differences in the minimum and optimum growth temperatures amongst the three isolates and the optimum growth temperature was 30 °C (Fig. 6). Research revealed that the highest growth rate of *M. paspali* occurred at 25–28 °C and *M. majus*, *M. seminicola* and *M. nivale* strains in Russia and Europe grew optimally at 20–25 °C (Doohan et al. 2003; Gagkaeva et al. 2020), while *M. nivale* from Slovakia grew better at temperatures below 20 °C (Hudec and Muchová 2010). Thus, the optimum growth temperature varies amongst *Microdochium* species.

A previous study showed that *P. herbarum* could initially induce leaf spots and blight on vetiver grass, causing round or irregular dark brown spots, which

are similar to the symptoms on *M. chrysopogonis* (Zhang et al. 2017). However, the symptoms on *M. chrysopogonis* were different from those on *P. herbarum* in the later period. Specifically, *P. herbarum* caused fusiform or irregular with red-dish-brown margins on the host plant, whereas *M. chrysopogonis* caused fish-eye necrotic haloes surrounding the spot lesions on leaves. Additionally, the disease incidences were different. *P. herbarum* affected 26% to 42% of plants, while *M. chrysopogonis* showed a 100% disease incidence. Given the high disease incidence associated with *M. chrysopogonis* and its induction of leaf spots on vetiver grass, as well as the identification of this new species, it is imperative to conduct future studies addressing the host spectrum, epidemic conditions, biological characteristics and distribution patterns of *M. chrysopogonis*.

The effectiveness of biofungicides, such as bacterial seed treatments using *Pseudomonas* and *Pantoea* in controlling diseases caused by *Microdochium*, has been established (Johansson et al. 2003). However, there remains substantial reliance on registered chemical fungicides. Currently, research on fungicide sensitivity within *Microdochium* mainly focuses on three species: *M. panattonianum*, *M. majus* and *M. nivale*. Six groups of fungicides, namely, MBCs, DMIs, QoIs, SDHIs, PPs, and dicarboximides, have been shown to have significant inhibitory activity (Kaneko and Ishii 2009; Aamlid et al. 2017, 2018; Matušinsky et al. 2017; Gagkaeva et al. 2022). In this study, consistent with previous findings, four DMI fungicides (difenoconazole, propiconazole, flusilazole and tebuconazole) and one MBC fungicide (carbendazim) exhibited significant inhibitory effects on the growth of *M. chrysopogonis*, with mean EC_{50} values of 0.077, 0.011, 0.004, 0.024 and 0.007 $\mu\text{g/ml}$, respectively (Table 4). However, dicarboximides (iprodione), which were effective against snow mould and *Microdochium* patch caused by *M. nivale* on turf-grass in previous studies (Gourlie and Hsiang 2017), showed ineffectiveness in this study, with a mean EC_{50} value of 193.031 $\mu\text{g/ml}$. Additionally, while the PP fungicide fludioxonil demonstrated good antifungal activity against *M. majus* in other research (Mao et al. 2023), the isolates in this study displayed only moderate sensitivity to fludioxonil, with an EC_{50} value of 4.525 $\mu\text{g/ml}$ and complete inhibition of mycelial growth required concentrations exceeding 100 $\mu\text{g/ml}$ (Table 4, Fig. 8). These variations in fungicide sensitivity could be attributed to genetic structural changes, introducing bias in chemical control efficacy (Matušinsky et al. 2019). Furthermore, the response of the same pathogen to fungicides can vary amongst regions. For example, the DMI fungicides, tebuconazole and metconazole, were reported to be ineffective against *M. nivale* in the Czech Republic and France (Ioos et al. 2005; Matušinsky et al. 2019). Similarly, *M. nivale* exhibited sensitivity to SDHI fungicides, including pydiflumetofen, fluxapyroxad and penthiopyrad, in vitro, but these fungicides were ineffective in providing acceptable control under field conditions in the USA (Hockemeyer and Koch 2022). These differences may be attributed to variations in environmental factors, such as temperature and humidity, as well as diverse biological characteristics, including epidemiology, fungicide sensitivity and aggressive nature of the pathogen (Abdelhalim et al. 2020). Overall, this study offers valuable insights into fungicide application strategies for effectively managing the disease. Further research is needed to analyse the influences of environmental variables and conduct field trials to validate the effects of DMI fungicides, ultimately enhancing the ability to successfully manage vetiver leaf tar spot disease.

Acknowledgements

We thank the Herbarium of the Chinese Academy of Forestry for helping with the preservation of plant specimens.

Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

Funding

This research was financially supported by the Guangdong Basic and Applied Basic Research Foundation (2020A1515110167), School-level Talents Project of Lingnan Normal University (ZL 2034), Natural Science Foundation of Guangdong Province, China (2023A1515011676) and Key Scientific Research Platform and Project of Guangdong Education Department (2021KCXTD054)

Author contributions

Xiang Lu, Xue-Li Niu and Wu Zhang carried out the investigation and sampling. Xiang Lu and Wu Zhang conducted the morphological and phylogenetic analysis. Xiang Lu and Wu Zhang carried out the pathogenicity test. Xiang Lu, Meng-Xian Mai, Wen-Hui Tan, Mu-Yan Zhang, Jie Xie and Yi Lu undertook the fungicide sensitivity experiment. Xiang Lu wrote, edited and reviewed the manuscript. Xiang Lu and Wu Zhang reviewed the manuscript and provided funding. All authors have read and agreed to the published version of the manuscript.

Author ORCIDs

Xiang Lu  <https://orcid.org/0000-0001-9582-1319>

Mengxian Mai  <https://orcid.org/0009-0001-3824-2895>

Wenhui Tan  <https://orcid.org/0009-0008-6054-2174>

Muyan Zhang  <https://orcid.org/0009-0005-8880-9780>

Jie Xie  <https://orcid.org/0009-0002-4200-0140>

Yi Lu  <https://orcid.org/0009-0003-0848-2390>

Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

References

- Aamlid TS, Espevig T, Tronsmo A (2017) Microbiological products for control of *Microdochium nivale* on golf greens. *Crop Science* 57(2): 559–566. <https://doi.org/10.2135/cropsci2016.05.0301>
- Aamlid TS, Niemeläinen O, Paaske K, Widmark D, Ruuttunen P, Kedonperä A (2018) Evaluation of a petroleum-derived spray oil for control of *Microdochium* patch and turf-grass spring performance on Nordic golf greens. *Agronomy Journal* 110(6): 2189–2197. <https://doi.org/10.2134/agronj2018.07.0475>

- Abdelhalim M, Brurberg MB, Hofgaard IS, Rognli OA, Tronsmo AM (2020) Pathogenicity, host specificity and genetic diversity in Norwegian isolates of *Microdochium nivale* and *Microdochium majus*. *European Journal of Plant Pathology* 156(3): 885–895. <https://doi.org/10.1007/s10658-020-01939-5>
- Alkan M, Özer G, İmren M, Özdemir F, Morgounov A, Dababat AA (2021) First report of *Fusarium culmorum* and *Microdochium bolleyi* causing root rot on *Triticale* in Kazakhstan. *Plant Disease* 105(7): e2015. <https://doi.org/10.1094/PDIS-12-20-2659-PDN>
- Araujo L, Paschoalino RS, Rodrigues FÁ (2016) Microscopic aspects of silicon-mediated rice resistance to leaf scald. *Phytopathology* 106(2): 132–141. <https://doi.org/10.1094/PHYTO-04-15-0109-R>
- Babu S, Rameshkumar S, Prakash M (2021) Vetiver-a Blessing to Coastal Ecosystem for an Integral Prosperity and Ecological Stability. *Coastal Agriculture and Climate Change*. CRC Press, Florida, 13 pp. <https://doi.org/10.1201/9781003245285-11>
- Braun U (1995) A Monograph of *Cercospora*, *Ramularia* and Allied Genera (Phytopathogenic Hyphomycetes). IHW Verlag, Eching, 2 pp.
- Chen XW, Wong JTF, Wang JJ, Wong MH (2021) Vetiver grass-microbe interactions for soil remediation. *Critical Reviews in Environmental Science and Technology* 51(9): 897–938. <https://doi.org/10.1080/10643389.2020.1738193>
- Crous PW, Slippers B, Wingfield MJ, Rheeder J, Marasas WFO, Philips AJL, Alves A, Burgess T, Barber P, Groenewald JZ (2006) Phylogenetic lineages in the Botryosphaeriaceae. *Studies in Mycology* 55: 235–253. <https://doi.org/10.3114/sim.55.1.235>
- Crous PW, Verkley GJM, Groenewald JZ, Samson RA (2009) *Fungal Biodiversity*. American Phytopathological Society, St. Paul, 269 pp.
- Crous PW, Wingfield MJ, Chooi Y-H, Gilchrist CLM, Lacey E, Pitt JI, Roets F, Swart WJ, Cano-Lira JF, Valenzuela-Lopez N, Hubka V, Shivas RG, Stchigel AM, Holdom DG, Jurjević Ž, Kachalkin AV, Lebel T, Lock C, Martín MP, Tan YP, Tomashevskaya MA, Vitelli JS, Baseia IG, Bhatt VK, Brandrud TE, De Souza JT, Dima B, Lacey HJ, Lombard L, Johnston PR, Morte A, Papp V, Rodríguez A, Rodríguez-Andrade E, Semwal KC, Tegart L, Abad ZG, Akulov A, Alvarado P, Alves A, Andrade JP, Arenas F, Asenjo C, Ballarà J, Barrett MD, Berná LM, Berraf-Tebbal A, Bianchinotti MV, Bransgrove K, Burgess TI, Carmo FS, Chávez R, Čmoková A, Dearnaley JDW, Santiago ALCMA, Freitas-Neto JF, Denman S, Douglas B, Dovana F, Eichmeier A, Esteve-Raventós F, Farid A, Fedosova AG, Ferisin G, Ferreira RJ, Ferrer A, Figueiredo CN, Figueiredo YF, Reinoso-Fuentealba CG, Garrido-Benavent I, Cañete-Gibas CF, Gil-Durán C, Glushakova AM, Gonçalves MFM, González M, Gorczak M, Gorton C, Guard FE, Guarnizo AL, Guarro J, Gutiérrez M, Hamal P, Hien LT, Hocking AD, Houbraken J, Hunter GC, Inácio CA, Jourdan M, Kapitonov VI, Kelly L, Khanh TN, Kislo K, Kiss L, Kiyashko A, Kolařík M, Kruse J, Kubátová A, Kučera V, Kučerová I, Kušan I, Lee HB, Levicán G, Lewis A, Liem NV, Liimatainen K, Lim HJ, Lyons MN, Maciá-Vicente JG, Magaña-Deñás V, Mahiques R, Malysheva EF, Marbach PAS, Marinho P, Matočec N, McTaggart AR, Mešić A, Morin L, Muñoz-Mohedano JM, Navarro-Ródenas A, Nicolli CP, Oliveira RL, Otsing E, Ovrebo CL, Pankratov TA, Paños A, Paz-Conde A, Pérez-Sierra A, Phosri C, Pintos Á, Pošta A, Prencipe S, Rubio E, Saitta A, Sales LS, Sanhueza L, Shuttleworth LA, Smith J, Smith ME, Spadaro D, Spetik M, Sochor M, Sochorová Z, Sousa JO, Suwannasai N, Tedersoo L, Thanh HM, Thao LD, Tkálčec Z, Vaghefi N, Venzhik AS, Verbeken A, Vizzini A, Voyron S, Wainhouse M, Whalley AJS, Wrzosek M, Zapata M, Zeil-Rolfe I, Groenewald JZ (2020) Fungal Planet description sheets: 1042–1111. *Persoonia* 44(1): 301–459. <https://doi.org/10.3767/persoonia.2020.44.11>
- Crous PW, Cowan DA, Maggs-Kölling G, Yilmaz N, Thangavel R, Wingfield MJ, Noorde-loos ME, Dima B, Brandrud TE, Jansen GM, Morozova OV, Vila J, Shivas RG, Tan YP,

- Bishop-Hurley S, Lacey E, Marney TS, Larsson E, Le Floch G, Lombard L, Nodet P, Hubka V, Alvarado P, Berraf-Tebbal A, Reyes JD, Delgado G, Eichmeier A, Jordal JB, Kachalkin AV, Kubátová A, Maciá-Vicente JG, Malysheva EF, Papp V, Rajeshkumar KC, Sharma A, Spetik M, Szabóová D, Tomashevskaya MA, Abad JA, Abad ZG, Alexandrova AV, Anand G, Arenas F, Ashtekar N, Balashov S, Bañares Á, Baroncelli R, Bera I, Biketova AYU, Blomquist CL, Boekhout T, Boertmann D, Bulyonkova TM, Burgess TI, Carnegie AJ, Cobo-Diaz JF, Corriol G, Cunningham JH, da Cruz MO, Damm U, Davoodian N, de A Santiago ALCM, Dearnaley J, de Freitas LWS, Dhileepan K, Dimitrov R, Di Piazza S, Fatima S, Fuljer F, Galera H, Ghosh A, Giraldo A, Glushakova AM, Gorczak M, Gouliamova DE, Gramaje D, Groenewald M, Gunsch CK, Gutiérrez A, Holdom D, Houbraken J, Ismailov AB, Istel Ł, Iturriaga T, Jeppson M, Jurjević Ž, Kalinina LB, Kapitonov VI, Kautmanová I, Khalid AN, Kiran M, Kiss L, Kovács Á, Kurose D, Kušan I, Lad S, Læssøe T, Lee HB, Luangsa-ard JJ, Lynch M, Mahamedi AE, Malysheva VF, Mateos A, Matočec N, Mešić A, Miller AN, Mongkolsamrit S, Moreno G, Morte A, Mostowfizadeh-Ghalamfarsa R, Naseer A, Navarro-Ródenas A, Nguyen TTT, Noisripoom W, Ntandu JE, Nuytinck J, Ostrý V, Pankratov TA, Pawłowska J, Pecenka J, Pham THG, Polhorský A, Pošta A, Raudabaugh DB, Reschke K, Rodríguez A, Romero M, Rooney-Latham S, Roux J, Sandoval-Denis M, Smith MTh, Steinrucken TV, Svetasheva TY, Tkalčec Z, van der Linde EJ, Vegte M, Vauras J, Verbeken A, Visagie CM, Vitelli JS, Volobuev SV, Weill A, Wrzosek M, Zmitrovich IV, Zvyagina EA, Groenewald JZ (2021a) Fungal Planet description sheets: 1182–1283. *Persoonia* 46: 313–528. <https://doi.org/10.3767/persoonia.2021.46.11>
- Crous PW, Osieck ER, Jurjević Ž, Boers J, Van Iperen AL, Starink-Willemse M, Dima B, Balashov S, Bulgakov TS, Johnston PR, Morozova OV, Pinruan U, Sommai S, Alvarado P, Decock CA, Lebel T, McMullan-Fisher S, Moreno G, Shivas RG, Zhao L, Abdollahzadeh J, Abrinbana M, Ageev DV, Akhmetova G, Alexandrova AV, Altés A, Amaral AGG, Angelini C, Antonín V, Arenas F, Asselman P, Badali F, Baghela A, Bañares A, Barreto RW, Baseia IG, Bellanger J-M, Berraf-Tebbal A, Biketova AYU, Bukharova NV, Burgess TI, Cabero J, Câmara MPS, Cano-Lira JF, Ceryngier P, Chávez R, Cowan DA, de Lima AF, Oliveira RL, Denman S, Dang QN, Dovana F, Duarte IG, Eichmeier A, Erhard A, Esteve-Raventós F, Fellin A, Ferisin G, Ferreira RJ, Ferrer A, Finy P, Gaya E, Geering ADW, Gil-Durán C, Glässnerová K, Glushakova AM, Gramaje D, Guard FE, Guarnizo AL, Haelewaters D, Halling RE, Hill R, Hirooka Y, Hubka V, Iliushin VA, Ivanova DD, Ivanushkina NE, Jangsantear P, Justo A, Kachalkin AV, Kato S, Khamsuntorn P, Kirtsideli IY, Knapp DG, Kochkina GA, Koukol O, Kovács GM, Kruse J, Kumar TKA, Kušan I, Læssøe T, Larsson E, Lebeuf R, Levicán G, Loizides M, Marinho P, Luangsa-ard JJ, Lukina EG, Magaña-Dueñas V, Maggs-Köling G, Malysheva EF, Malysheva VF, Martín B, Martín MP, Matočec N, McTaggart AR, Mehrabi-Koushki M, Mešić A, Miller AN, Mironova P, Moreau P-A, Morte A, Müller K, Nagy LG, Nanu S, Navarro-Ródenas A, Nel WJ, Nguyen TH, Nóbrega TF, Noordeloos ME, Olariaga I, Overton BE, Ozerskaya SM, Palani P, Pancorbo F, Papp V, Pawłowska J, Pham TQ, Phosri C, Popov ES, Portugal A, Pošta A, Reschke K, Reul M, Ricci GM, Rodríguez A, Romanowski J, Ruchikachorn N, Saar I, Safi A, Sakolrak B, Salzmann F, Sandoval-Denis M, Sangwichein E, Sanhueza L, Sato T, Sastoque A, Senn-Irlet B, Shibata A, Siepe K, Somrithipol S, Spetik M, Sridhar P, Stchigel AM, Stuskova K, Suwannasai N, Tan YP, Thangavel R, Tiago I, Tiwari S, Tkalčec Z, Tomashevskaya MA, Tonegawa C, Tran HX, Tran NT, Trovão J, Trubitsyn VE, Van Wyk J, Vieira WAS, Vila J, Visagie CM, Vizzini A, Volobuev SV, Vu DT, Wangsawat N, Yaguchi T, Ercole E, Ferreira BW, de Souza AP, Vieira BS, Groenewald JZ (2021b) Fungal Planet description sheets: 1284–1382. *Persoonia* 47: 178–374. <https://doi.org/10.3767/persoonia.2021.47.06>

- David A, Wang F, Sun X, Li H, Lin J, Li P, Deng G (2019) Chemical composition, antioxidant, and antimicrobial activities of *Vetiveria zizanioides* (L.) nash essential oil extracted by carbon dioxide expanded ethanol. *Molecules* (Basel, Switzerland) 24(10): 1897. <https://doi.org/10.3390/molecules24101897>
- Debieu D, Bach J, Arnold A, Brousset S, Gredt M, Taton M, Rahier A, Malosse C, Leroux P (2000) Inhibition of ergosterol biosynthesis by morpholine, piperidine, and spiroketalamine fungicides in *Microdochium nivale*: Effect on sterol composition and sterol $\Delta 8 \rightarrow \Delta 7$ -isomerase activity. *Pesticide Biochemistry and Physiology* 67(2): 85–94. <https://doi.org/10.1006/pest.2000.2485>
- Dirchwolf PM, Moschini RC, Gutiérrez SA, Carmona MA (2023) Modelling of the effects of environmental factors on rice grain discoloration incidence in Corrientes province, Argentina. *Journal of Phytopathology* 171(1): 12–22. <https://doi.org/10.1111/jph.13150>
- Doohan FM, Brennan J, Cooke BM (2003) *Epidemiology of Mycotoxin Producing Fungi*. Springer, Dordrecht, 129 pp.
- Gagkaeva TY, Orina AS, Gavrilova OP, Gogina NN (2020) Evidence of *Microdochium* Fungi associated with cereal grains in Russia. *Microorganisms* 8(3): e340. <https://doi.org/10.3390/microorganisms8030340>
- Gagkaeva T, Orina A, Gavrilova O, Usoltseva M, Crouch JA, Normann K, Entwistle K, Torp T, Espevig T (2022) In vitro fungicide sensitivity of *Clavireedia*, *Fusarium*, and *Microdochium* isolates from grasses. *International Turfgrass Society Research Journal* 14(1): 972–980. <https://doi.org/10.1002/its2.139>
- Galea VJ, Price TV, Sutton BC (1986) Taxonomy and biology of the lettuce anthracnose fungus. *Transactions of the British Mycological Society* 86(4): 619–628. [https://doi.org/10.1016/S0007-1536\(86\)80065-1](https://doi.org/10.1016/S0007-1536(86)80065-1)
- Glynn NC, Hare MC, Edwards SG (2008) Fungicide seed treatment efficacy against *Microdochium nivale* and *M. majus* in vitro and in vivo. *Pest Management Science* 64(8): 793–799. <https://doi.org/10.1002/ps.1558>
- Gourlie R, Hsiang T (2017) Resistance to dicarboximide fungicides in a Canadian population of *Microdochium nivale*. *International Turfgrass Society Research Journal* 13(1): 133–138. <https://doi.org/10.2134/itsrj2016.06.0497>
- Hernández-Restrepo M, Groenewald JZ, Crous PW (2016) Taxonomic and phylogenetic re-evaluation of *Microdochium*, *Monographella* and *Idriella*. *Persoonia* 36(1): 57–82. <https://doi.org/10.3767/003158516X688676>
- Hockemeyer K, Koch PL (2022) Field evaluations and in vitro sensitivity of *Microdochium nivale* to succinate dehydrogenase (SDHI) fungicides. *International Turfgrass Society Research Journal* 14(1): 951–957. <https://doi.org/10.1002/its2.48>
- Hong SK, Kim WG, Choi HW, Lee SY (2008) Identification of *Microdochium bolleyi* associated with basal rot of creeping bent grass in Korea. *Mycobiology* 36(2): 77–80. <https://doi.org/10.4489/MYCO.2008.36.2.077>
- Hudec K, Muchová D (2010) Influence of temperature and species origin on *Fusarium* spp. and *Microdochium nivale* pathogenicity to wheat seedlings. *Plant Protection Science* 46(2): 59–65. <https://doi.org/10.17221/12/2009-PPS>
- Ioos R, Belhadj A, Menez M, Faure A (2005) The effects of fungicides on *Fusarium* spp. and *Microdochium nivale* and their associated trichothecene mycotoxins in French naturally-infected cereal grains. *Crop Protection* 24(10): 894–902. <https://doi.org/10.1016/j.cropro.2005.01.014>
- Jewell LE, Hsiang T (2013) Multigene differences between *Microdochium nivale* and *Microdochium majus*. *Botany* 91(2): 99–106. <https://doi.org/10.1139/cjb-2012-0178>

- Jia X, Zheng X, Damm U, Lu X, Niu XL, Feng Y, Zhang W (2023 [June 04]) *Stenocarpella chrysopogonis*, a new species causing leaf streak disease of *Chrysopogon zizanioides* in southern China. Plant Disease PDIS-02-23-0383-SR. <https://doi.org/10.1094/PDIS-02-23-0383-SR>
- Johansson PM, Johnsson L, Gerhardson B (2003) Suppression of wheat-seedling diseases caused by *Fusarium culmorum* and *Microdochium nivale* using bacterial seed treatment. Plant Pathology 52(2): 219–227. <https://doi.org/10.1046/j.1365-3059.2003.00815.x>
- Kaneko I, Ishii H (2009) Effect of azoxystrobin on activities of antioxidant enzymes and alternative oxidase in wheat head blight pathogens *Fusarium graminearum* and *Microdochium nivale*. Journal of General Plant Pathology 75(5): 388–398. <https://doi.org/10.1007/s10327-009-0178-9>
- Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. Molecular Biology and Evolution 30(4): 772–780. <https://doi.org/10.1093/molbev/mst010>
- Koch PL, Stier JC, Kerns JP (2015) Snow cover has variable effects on persistence of fungicides and their suppression of microdochium patch on amenity turfgrass. Plant Pathology 64(6): 1417–1428. <https://doi.org/10.1111/ppa.12379>
- Kumar S, Stecher G, Tamura K (2016) MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular Biology and Evolution 33(7): 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Liang J, Li G, Zhao M, Cai L (2019) A new leaf blight disease of turfgrasses caused by *Microdochium poae*, sp. nov. Mycologia 111(2): 265–273. <https://doi.org/10.1080/00275514.2019.1569417>
- Lin JM, Liu JX, Niu XL, Zhang W (2019) Root and basal stem rot caused by *Gaeumannomyces graminicola* on *Vetiveria zizanioides* in southern China. Plant Disease 103(8): e2128. <https://doi.org/10.1094/PDIS-12-18-2161-PDN>
- Liu YJ, Whelen S, Hall BD (1999) Phylogenetic relationships among ascomycetes: Evidence from an RNA polymerase II subunit. Molecular Biology and Evolution 16(12): 1799–1808. <https://doi.org/10.1093/oxfordjournals.molbev.a026092>
- Mao Y, Wu J, Song W, Zhao B, Zhao H, Cai Y, Wang J-X, Zhou M, Duan Y (2023) Occurrence and chemical control strategy of wheat brown foot rot caused by *Microdochium majus*. Plant Disease 107(11): 3523–3530. <https://doi.org/10.1094/PDIS-02-23-0392-RE>
- Matušinsky P, Svačinová I, Jonavičienė A, Tvarůžek L (2017) Long-term dynamics of causative agents of stem base diseases in winter wheat and reaction of Czech *Oculimacula* spp. and *Microdochium* spp. populations to prochloraz. European Journal of Plant Pathology 148(1): 199–206. <https://doi.org/10.1007/s10658-016-1082-8>
- Matušinsky P, Svobodová LL, Svačinová I, Havis N, Hess M, Tvarůžek L (2019) Population genetic structure of *Microdochium majus* and *Microdochium nivale* associated with foot rot of cereals in the Czech Republic and adaptation to penthiopyrad. European Journal of Plant Pathology 155(1): 1–12. <https://doi.org/10.1007/s10658-019-01737-8>
- Maurya R, Yadav P, Krishna R, Kulkarni P, Rastogi S, Mohapatra S, Srivastava S, Akhtar MQ, Shukla AK, Chauhan HS, Verma RK, Chanotiya CS, Shasany AK (2023) Transcriptome and metabolome analysis of sesquiterpene diversity in Indian vetiver (*Chrysopogon zizanioides* L. Roberty). Industrial Crops and Products 200: e116798. <https://doi.org/10.1016/j.indcrop.2023.116798>
- Moon UR, Durge AA, Kumar M (2020) Value addition of khus (*Vetiveria zizanioides* L. Nash) root extracts by elicitor and heat treatment. Industrial Crops and Products 144: e112037. <https://doi.org/10.1016/j.indcrop.2019.112037>

- Nylander JAA (2004) MrModeltest 2.0. MrModeltest Version 2. Uppsala University, Uppsala.
- Polizzi G, Aiello D, Vitale A, Giuffrida F, Groenewald JZ, Crous PW (2009) First report of shoot blight, canker, and gummosis caused by *Neoscytalidium dimidiatum* on citrus in Italy. *Plant Disease* 93(11): 1215–1215. <https://doi.org/10.1094/PDIS-93-11-1215A>
- Rambaut A (2018) FigTree v.1.4.4. University of Edinburgh, Edinburgh.
- Rayner RW (1970) A Mycological Colour Chart. Commonwealth Mycological Institute & British Mycological Society, Kew, 34 pp.
- Ren R, Yang X, Ray RV (2015) Comparative aggressiveness of *Microdochium nivale* and *M. majus* and evaluation of screening methods for *Fusarium* seedling blight resistance in wheat cultivars. *European Journal of Plant Pathology* 141(2): 281–294. <https://doi.org/10.1007/s10658-014-0541-3>
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Stewart S, Rodríguez M, Mattos N, Abreo E, Stewart S, Rodríguez M, Mattos N, Abreo E (2019) First report of zonate leaf spot in sorghum caused by *Microdochium sorghi* in Uruguay. *Agrociencia* 23(2): 94–98. <https://doi.org/10.31285/AGRO.23.74>
- Swofford D (2002) PAUP*. phylogenetic analysis using parsimony (*and other methods). version 4.0b10. Sinauer Associates, Sunderland.
- Sydow H (1924) Mycotheca germanica. Fasc. XLII–XLV (No. 20512250). *Annales Mycologici* 22: 257–268.
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172(8): 4238–4246. <https://doi.org/10.1128/jb.172.8.4238-4246.1990>
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and Direct Sequencing of Fungal Ribosomal RNA Genes for Phylogenetics. *PCR Protocols: a Guide to Methods and Applications*. Academic Press, San Diego, 482 pp. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>
- Wicks TJ, Hall B, Pezzaniti P (1994) Fungicidal control of anthracnose (*Microdochium panattonianum*) on lettuce. *Australian Journal of Experimental Agriculture* 34(2): 277–283. <https://doi.org/10.1071/EA9940277>
- Yin LF, Du SF, Chaisiri C, Cheewangkoon R, Luo CX (2019) Phylogenetic analysis and fungicide baseline sensitivities of *Monilia mumeicola* in China. *Plant Disease* 103(9): 2231–2236. <https://doi.org/10.1094/PDIS-11-18-1953-RE>
- Zhang W, Nan Z, Tian P, Hu M, Gao Z, Li M, Liu G (2015) *Microdochium paspali*, a new species causing seashore paspalum disease in southern China. *Mycologia* 107(1): 80–89. <https://doi.org/10.3852/14-119>
- Zhang W, Liu JX, Huo PH (2017) *Phoma herbarum* causes leaf spots and blight on Vetiver grass (*Vetiveria zizanioides* L.) in southern China. *Plant Disease* 101(10): e1823. <https://doi.org/10.1094/PDIS-04-17-0519-PDN>

Supplementary material 1

Phylogenetic tree inferred from a maximum likelihood analysis based on a combined alignment of ITS sequences of 97 isolates of the *Microdochium* sp.

Authors: Xiang Lu, Mengxian Mai, Wenhui Tan, Muyan Zhang, Jie Xie, Yi Lu, Xue Li Niu, Wu Zhang

Data type: tif

Explanation note: Bootstrap support values obtained with ML above 70% and Bayesian (BI) posterior probability values above 0.90 are shown at the nodes (BI/ML). The designated outgroup taxa are *Thamnomycetes dendroidea* CBS 123578. Numbers of ex-type strains are emphasized with an asterisk and species are delimited with shaded blocks. Isolates of *M. chrysopogonis* are indicated with lighter text.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mycokeys.100.112128.suppl1>

Supplementary material 2

Phylogenetic tree inferred from a maximum likelihood analysis based on a combined alignment of LSU sequences of 72 isolates of the *Microdochium* sp.

Authors: Xiang Lu, Mengxian Mai, Wenhui Tan, Muyan Zhang, Jie Xie, Yi Lu, Xue Li Niu, Wu Zhang

Data type: tif

Explanation note: Bootstrap support values obtained with ML above 70% and Bayesian (BI) posterior probability values above 0.90 are shown at the nodes (BI/ML). The designated outgroup taxa are *Thamnomycetes dendroidea* CBS 123578. Numbers of ex-type strains are emphasized with an asterisk and species are delimited with shaded blocks. Isolates of *M. chrysopogonis* are indicated with lighter text.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mycokeys.100.112128.suppl2>

Supplementary material 3

Phylogenetic tree inferred from a maximum likelihood analysis based on a combined alignment of *rpb2* sequences of 71 isolates of the *Microdochium* sp.

Authors: Xiang Lu, Mengxian Mai, Wenhui Tan, Muyan Zhang, Jie Xie, Yi Lu, Xue Li Niu, Wu Zhang

Data type: tif

Explanation note: Bootstrap support values obtained with ML above 70% and Bayesian (BI) posterior probability values above 0.90 are shown at the nodes (BI/ML). The designated outgroup taxa are *Thamnomycetes dendroidea* CBS 123578. Numbers of ex-type strains are emphasized with an asterisk and species are delimited with shaded blocks. Isolates of *M. chrysopogonis* are indicated with lighter text.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mycokeys.100.112128.suppl3>

Supplementary material 4

Phylogenetic tree inferred from a maximum likelihood analysis based on a combined alignment of *tub2* sequences of 80 isolates of the *Microdochium* sp.

Authors: Xiang Lu, Mengxian Mai, Wenhui Tan, Muyan Zhang, Jie Xie, Yi Lu, Xue Li Niu, Wu Zhang

Data type: tif

Explanation note: Bootstrap support values obtained with ML above 70% and Bayesian (BI) posterior probability values above 0.90 are shown at the nodes (BI/ML). The designated outgroup taxa are *Thamnomycetes dendroidea* CBS 123578. Numbers of ex-type strains are emphasized with an asterisk and species are delimited with shaded blocks. Isolates of *M. chrysopogonis* are indicated with lighter text.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mycokeys.100.112128.suppl4>